Asian Journal of Soil Science and Plant Nutrition

4(1): 1-16, 2019; Article no.AJSSPN.46028 ISSN: 2456-9682

Influence of Foliar Application with Plant Aqueous Extracts on Growth, Yield and Chemical Constituents of Chamomile

Yasmin M. R. Abdellatif^{1*} and Hemmat, A. Ibrahim²

1 Department of Agricultural Botany, Faculty of Agriculture, Ain Shams University, Cairo 11241, Egypt. ² Department of Agricultural Biochemistry, Faculty of Agriculture, Ain Shams University, Egypt.

Authors' contributions

This work was carried out in collaboration between both authors. Authors YMRA and HAI designed the study, wrote the protocol, cultured the experiment, performed the chemical and statistical analyses and prepared the manuscript. Both authors read and approved the final manuscript.

Article Information

DOI: 10.9734/AJSSPN/2019/46028 *Editor(s):* (1) Dr. Kosev Valentin, Associate Professor, Institute of Forage Crops, Pleven, Bulgaria. (2) Dr. Tancredo Souza, Professor, Department of Life Sciences, Centre for Functional Ecology, University of Coimbra, Portugal. (3) Dr. Abhishek Naik, Department of Area Manager in Technology Development, Vegetable Crops Organization-United Phosphorus Limited, India. *Reviewers:* (1) Ernest Eteng, Michael Okpara IUniversity of Agriculture, Nigeria. (2) Florin Sala, Banat University of Agricultural Sciences and Veterinary Medicine "King Michael I of Romania" from Timisoara, Romania. Complete Peer review History: http://www.sdiarticle3.com/review-history/46028

> *Received 12 November 2018 Accepted 23 January 2019 Published 28 January 2019*

Original Research Article

ABSTRACT

The main target of sustainable agriculture including organic farming is to use natural compounds such as plant aqueous extracts to elevate plant growth and productivity. The subject of the present study is to determine the plant growth and inflorescences production, some biochemical constituents of shoot and inflorescences and antioxidative activities of essential oil obtained from chamomile plants exogenously sprayed with aqueous extracts of dried roselle calyces, turmeric rhizomes, safflower flowers and red beet roots. A pot experiment was conducted during the two successive seasons of 2016/2017 and 2017/2018 in the open field of Experimental Farm of Agricultural Botany Department, Faculty of Agriculture, Ain Shams university, Qalyubia, Egypt. Transplants of chamomile, 45 days old, were separately sprayed after 15 days from transplanting by the four different aqueous extracts and distilled water was used as a control. Generally, spraying with tested plant aqueous extracts on chamomile plants caused high efficiency in growth promotion, inflorescences and essential oil production. Red beet and safflower extracts gave the

**Corresponding author: Email: dr_yasminmarzouk@agr.asu.edu.eg;*

highest number of branches and inflorescences per plant. Chlorophyll a, b, carotenoids, reducing sugars and amino acids were increased in chamomile shoots when red beet and safflower extracts were sprayed while flavonoids and phenolic compounds were significantly decreased in comparing with roselle and turmeric extract treatments. Different concentrations of essential oil and inflorescences ethanolic extracts obtained from chamomile plants treated with safflower and red beet extracts showed the highest scavenging activities on DPPH radical and lowest IC_{50} values. Finally, it could be concluded that application of plant aqueous extracts considered as alternative method to chemical compounds which achieved sustainability of organic farming.

Keywords: Chamomile (Matricaria chamomilla L.); Roselle extract; turmeric extract; safflower extract; red beet extract; essential oil; DPPH radical; IC50.

1. INTRODUCTION

Recently, public health and environmental safety encourage the use of plant extracts for improving growth, chemical composition and productivity of plants especially medicinal plants. Chamomile is one the important medicinal and aromatic plant belong to *Asteraceae* family, has a sweet, grassy and lovely fruity aroma. It has many medicinal uses due to its calming, carminative and spasmolytic properties, antimicrobial and antiinflammatory effects [1]. The main bioactive constituents of chamomile essential oil are αbisabolol, bisabolol oxide A, bisabolol oxide B, bisabolone oxide, α-pinene, β-pinene, chamazulene, camphene, myrcene, sabinene, 1,8-cineole y-terpinene, caryophyllene, propyl angelate and butyl angelate. Also, flavone glucosides (apigenin 7-O-glucoside and various acylated derivatives of apigenin 7-O-glucoside) and flavonols (luteolin glucosides, quercetin and isohamnetin glucosides) were identified in chamomile [2].

However, plant extracts have recently become more common applications in modern agricultural production, among these substances are roselle, turmeric, safflower and red beet aqueous extracts.

Roselle (*Hibiscus sabdariffa* L.) is a tropical shrub with red, dark red or green inflated edible calyces belongs to family *Malvaceae* [3,4]. The calyces have been found to be rich in vitamin C and other antioxidants such as flavonoids [5] and minerals [6].

Turmeric, *Curcuma longa* L*.* (*Zingiberaceae*) rhizome commonly used as a spice [7]. The higher content of turmeric from amino acids, potassium, vitamins, antioxidants and plant pigments as curcumin and volatile oils encourage to undertake many attempts for using its aqueous extract as a stimulator for plant growth [8].

Safflower (*Carthamus tinctorius* L.) is world's oldest crop belonging to family *Compositae* which contains water soluble yellow dye (carthamidin), it has been used traditionally as an annual oil seed crop, medicinal herb and natural dye source for coloring food and textile [9].

Red beet (*Beta vulgaris* L.) is an herbaceous biennial crop belonging to family *Chenopodiaceae*, deep red-colored beet bulbs are the most common consumed for human [10]. Beet roots are rich in valuable bioactive compounds such as ß-cyanines [11], glycine betaine [12], saponins [13], carotenoids [14], betanin, polyphenols and flavonoids [15]. It considered as one of the most potent vegetables contains antioxidants due to the presence of ßcyanins which are a group of compounds exhibiting antioxidant and radical-scavenging activities [16].

The purpose of this study is to investigate the effect of roselle, turmeric, safflower and red beet aqueous extracts as foliar sprayers on growth characters and yield production of chamomile plants and evaluate the stimulation of these extracts on the essential oil yield and the antioxidant potential of treated chamomile plants.

2. MATERIALS AND METHODS

2.1 Plant Material

Seeds of chamomile (*Matricaria chamomilla* L.) were kindly produced from the Aromatic and Medicinal Plant Research Institute, ARC, Ministry of Agriculture, Egypt.

2.2 Preparation of Plant Aqueous Extracts

Calyces of roselle (*Hibiscus sabdariffa* L.), rhizomes of turmeric (*Curcuma longa* L*.*), flowers of safflower (*Carthamus tinctorius* L.) were produced from Aromatic and Medicinal Plant Research Institute and roots of red beet (*Beta vulgaris* L.) were produced from Horticulture Research Institute, ARC, Ministry of Agriculture, Egypt, then dried, grinded and macerated 2 g powder in 100 ml of distilled water and soaked for 24 h then filtered and used freshly.

2.3 Chemical Analysis of Plant Aqueous Extracts

The pH values, titratable acidity, soluble phenolic compounds, total flavonoids, anthocyanin, reducing sugars, free amino acids and N, P, K percentages were determined in the previous aqueous extracts.

Total titratable acidity (mg citric acid 100 mg $^{-1}$ d.wt.) of plant aqueous extracts was determined according to A.O.A.C. [17].

Total soluble phenols, flavonoids, reducing sugars and free amino acids were extracted from chamomile shoots according to Ackerson [18] using 80% ethanol.

Soluble phenolic compounds were estimated as (g $100 g^{-1}$ d.wt.) by the method of Folin-Ciocalteu as described by Shahidi and Naczk [19] using gallic acid as a standard.

Total flavonoids concentration was determined as (g 100 g^{-1} d.wt.) by the aluminum chloride colorimetric assay according to Marinova et al. [20] using quercetin as a standard.

Anthocyanins concentration was colorimetrically proceeded as g 100 g^{-1} d.wt. according to Du and Francis [21].

Reducing sugars were determined colorimeterically $(g 100g⁻¹ d.wt.)$ by using 3,5 dinitrosalsylic acid according to Miller [22] using glucose as a standard.

Free amino acids were estimated colorimetrically (g $100g^{-1}$ d.wt.) by using ninhydrin according to Jayeraman [23] using glycine as a standard.

Plant aqueous extracts were digested by using $H₂SO₄$ and $H₂O₂$ according to the method described by Piper [24] to determine N, P and K percentages according to the method described by Black et al. [25] and Wilde et al. [26].

2.4 Experimental Set up

Pot experiment was carried out under open field condition in the Farm of Agricultural Botany Department, Faculty of Agriculture, Ain Shams University, Shoubra El Kheima, Qalyubia, Egypt, (30°06′42″ N 31°14′46″ E) during the two successive winter seasons of 2016/2017 and 2017/2018. Chamomile seeds were sown on 15th September in nursery beds. On 1st November of both seasons, when the grown seedlings reached about 10-15 cm in length, ten transplants were transferred in plastic pots (30 x 40 cm) filled with clay loamy/sand (3:1 v/v) soil (Table 1). The plants were thinned out into three uniform transplants per each pot after 10 days from transplanting (DAT). Three replicates for each treatment, three pots/replicate, were arranged in a randomized complete block design. The pots were regularly irrigated with tap water when plants needed. Each seedling was sprayed with 30 ml of the four previous prepared fresh aqueous extracts (2% w/v) and tap water was used as control. The volume of extracts was consequently increased with increasing plant growth. The foliar applications were applied four times, the first one was carried out 15 days after transplanting and others were applied with 2 weeks intervals. Tween 20 at 0.1% was used as a wetting agent. All agricultural practices were done as the recommendations of Ministry of Agriculture.

2.5 Chamomile Plant Samples

Three vegetative plant replicates were randomly taken from each treatment at 45 days after transplanting, after the third spray treatment, for the chemical analyses, *i.e.* chlorophyll a (Chl a), chlorophyll b (Chl b), carotenoids, flavonoids, total soluble phenolic compounds, reducing sugars (RS) and amino acids (AA).

Table 1. Mechanical and chemical analyses of the experimental soil

Another two samples (at 45 and 120 days after transplanting) were randomly collected to measure plant height (cm), number of branches plant⁻¹, shoot fresh and dry weights plant⁻¹ (g). Each sample was contained three plants from each treatment.

Harvesting of inflorescences started at the last week of January until the middle of April for the both seasons. The picking of the inflorescences was done continuously when the ray flowers were in mood. Three plants were used to determine the total number of inflorescences plant⁻¹, total inflorescences fresh and dry weights plant⁻¹ (g) .

Total soluble phenolic compounds, flavonoids and carotenoids were determined in chamomile inflorescences. The essential oil yield plant $⁻¹$, its</sup> chemical components and scavenging activity on DPPH radical in both oil and inflorescences ethanolic extract were estimated.

2.6 Chemical Analyses of Chamomile Samples

Chlorophylls a & b (Chl a, Chl b) and carotenoids concentrations were extracted and assayed as the procedure of Costache et al. [27]. Chl a, b and carotenoids were expressed in shoots as mg g^{-1} f.wt. whereas carotenoids were estimated in inflorescences as mg 100 g^{-1} d.wt.

Soluble phenolic compounds, total flavonoids, reducing sugars and free amino acids concentrations were determined as mentioned before.

Inflorescences essential oil was distilled using a micro distilling apparatus and oil volume was measured as ml of oil $100g⁻¹$ d.wt. inflorescences according to Guenther [28].

Chemical components of the chamomile essential oil were determined by Gas liquid chromatography-mass spectrometer (GLC-MS). Chromatographic analysis of essential oil using GC-MS was performed (Agilent Technologies 7890 GC system combined with 5977, A Mass Selective Detector) in National Research Center, El Dokki, Egypt.

Ethanolic solutions of chamomile essential oil (2, 4, 6, 8 and 10 μ g ml⁻¹) and inflorescences ethanolic extract (0.66, 1.33, 2.00, 2.66 and 3.33 mg ml⁻¹) were prepared. The percent of scavenging activity of the different concentrations

of both extracted essential oil and inflorescences ethanolic extracts on DPPH radical were evaluated by measuring from the bleaching of the violet colored ethanolic solution of DPPH according to Gulluce et al. [29]. This spectrophotometric assay uses the stable radical 2, 2′- diphenyl-1-picryl hydrazyl (DPPH) as a reagent. Inhibition of free radical DPPH calculated according to the following equation:

Scavenging activity (%)= [(Acontrol - Asample)/Acontrol]x 100

The concentrations of the chamomile essential oil (μ g ml⁻¹) and the inflorescences ethanolic extract (mg ml^{-1}) that needed to inhibit 50% of the initial DPPH concentration (IC_{50}) was calculated for each treatment.

2.7 Statistical Analysis

All experiment data was analyzed by analysis of variance (ANOVA) using the General Linear Model procedure of CoStat. Significance between means was tested by "F" test and the value of LSD (p≤0.05) was calculated [30]. Significant differences of means at *P* ≤ 0.05 were compared by different letters as described by Duncan test of Gomez and Gomez [31]. Results expressed as mean ± standard deviation (SD).

3. RESULTS

3.1 Chemical Analyses of Plant Aqueous Extracts

Data in Table 2 showed significant differences (P≤0.05) in the physical-biochemical analyses of all plant aqueous extracts. All extracts had acidity pH values. Roselle aqueous extract had the lowest pH value (2.62) followed by red beet extract (4.42), safflower extract (4.77) and turmeric extract which had the highest pH value (5.84). In contrast, titratable acidity % (TA%) showed the opposite trend where roselle extract contained the highest TA% (15.66%) while turmeric extract had the lowest percent (0.96%).

Safflower aqueous extract contained the highest concentrations of total soluble phenols (2.55%), reducing sugars $(15.47%)$ and P $(0.80%)$ in addition to its high concentration of amino acids. whereas, the highest concentration of free amino acids (1.17%), flavonoids (8.46%) and K (7.85%) were recorded in red beet extract. Roselle extract was found to contain the highest concentration of N compared to other extracts (Table 2). It seemed also that turmeric extract was low in

most of biochemical estimates that were appreciated. As for anthocyanin, roselle and red beet extracts contained (0.29% and 0.09%), respectively while turmeric and safflower extracts recorded anthocyanin free aqueous extracts.

3.2 Effects of Foliar Application with Plant Aqueous Extracts on Vegetative Growth Parameters and Inflorescences Yield of Chamomile Plants

Significant differences in growth parameters of chamomile shoot was detected by foliar applications of plant aqueous extracts at 45 days

(Table 3) and 120 days (Table 4) after transplanting during the two seasons 2016/2017 and 2017/2018.

Obtained results in Table 3 revealed that plant growth remarkably response to the different exogenous spray extracts compared to control. Chamomile plants sprayed by roselle or turmeric extracts showed an increase in plant height more than plants treated with safflower or red beet extracts, on the other side, safflower or red beet extracts enhanced the number of branches/plant and shoot f.wt. and d.wt. at 45 days after transplanting (DAT) in the two seasons.

Table 2. Some physical properties and biochemical constituents of roselle, turmeric, safflower and red beet aqueous extracts

Physical properties	Plant aqueous extracts					
and biochemical analyses	Roselle extract	Turmeric extract	Safflower extract	Red beet extract		
рH	2.62° ±0.01	5.84^{a} ±0.01	4.77° ±0.01	4.42° ±0.01		
Titratable acidity	$15.66^a \pm 0.57$	0.96° ±0.005	2.55° ±0.005	$4.76^{\rm b}$ ±0.05		
(mg citric acid 100 g^{-1} d.wt.)						
Soluble phenols %	$1.77^{\rm b}$ ±0.15	0.14° ±0.002	2.55° ±0.02	0.32° ±0.015		
Flavonoids %	$2.44^b \pm 0.08$	0.29° ±0.001	1.76° ±0.01	$8.46^a \pm 0.007$		
Anthocyanin	$0.29^a \pm 0.001$	0.00	0.00	$0.09^{\rm b}$ ±0.001		
mg $100g^{-1}$ d.wt.						
Reducing sugars%	6.92° ±0.01	1.18° ±0.01	15.47° ±0.02	12.78° ±0.01		
Free amino acids %	0.14° ±0.006	0.15° ±0.002	$1.09^{\rm b}$ ±0.02	$1.17^a \pm 0.04$		
$N\%$	40.92° ±1.57	11.52° ±0.37	$20.55^b \pm 0.79$	$21.39^b \pm 0.95$		
P%	$0.59^{\rm b}$ ±0.07	$0.37^d \pm 0.001$	$0.80^a \pm 0.001$	0.48° ±0.001		
K%	$7.57^b \pm 0.01$	5.65° ±0.01	6.18° ±0.01	7.85° ±0.01		

Data were presented as mean ± SD (n=3). Means in the same row with different letters are significantly different at P≤0.05

Table 3. Growth parameters "at 45 DAT" of chamomile plants sprayed with four different aqueous extracts during 2016/2017 and 2017/2018 successive seasons

Data were presented as mean ± SD (n=3). Means in the same row with different letters are significantly different at P≤0.05

Abdellatif and Ibrahim; AJSSPN, 4(1): 1-16, 2019; Article no.AJSSPN.46028

Moreover, foliar spray with turmeric extract influenced the highest chamomile shoot height (70.50 and 73.50 cm) followed by roselle water extract (66.60 and 66.50 cm), at120 DAT of the two seasons, respectively. Both safflower and red beet extracts increased number of branches and shoot f.wt. and d.wt. comparing to control plants. Number of branches/plant increased about 3-4 folds more than the control plants with all different aqueous extracts at both seasons of experiment (Table 4).

In the same trend foliar applied roselle, turmeric, red beet and safflower extracts gradually increased total no. of inflorescences, inflorescences fresh and dry weights per plant, these increases reached the significant level when compared to control (Table 5).

3.3 Chemical Constituents of Chamomile Shoot Influenced with Foliar Applications of Plant Aqueous Extracts

Tables (6 and 7) showed significant increase (P≤0.05), in most cases, in the tested chemical constituents of chamomile shoots sprayed with the four plant aqueous extracts.

Table 4. Growth parameters "at 120 DAT" of chamomile plants sprayed with four different aqueous extracts during 2016/2017 and 2017/2018 successive seasons

Data were presented as mean ± SD (n=3). Means in the same row with different letters are significantly different at P≤0.05

Table 5. Inflorescences yield "at 150 DAT" of chamomile per plant sprayed with four different aqueous extracts during 2016/2017 and 2017/2018 successive seasons

Data were presented as mean ± SD (n=3). Means in the same row with different letters are significantly different at P≤0.05

3.3.1 Chl a, Chl b and carotenoids

Foliar applications with red beet and safflower extracts induced markedly increase in Chl a (0.94 and 0.92), Chl b (0.35 and 0.34) and carotenoids (0.25 and 0.24) mg/g f.wt. in chamomile shoots in comparison with untreated plants which showed the lowest pigments concentration (0.79, 0.27 and 0.22 mg/g f.wt) respectively, at the first season (Table 6). Also, the same behavior was detected at the second one (Table 7).

3.3.2 Flavonoids and soluble phenolic compounds

Tables (6 and 7) showed also that exogenous applied turmeric extract elevated the total flavonoid concentrations (115.40 and 101.20 mg 100g-1 f.wt.) and soluble phenolic compounds concentrations (114.07 and 103.76 mg $100g^{-1}$

f.wt.) more than control plants in chamomile shoots during the two successive seasons, respectively. Flavonoids concentration was also increased about 2 - 3.5 times than control when plants treated with the other plant aqueous extracts. On the other hand, soluble phenols were decreased due to spraying with safflowers and red beet extracts at the both seasons when compared with control.

3.3.3 Reducing sugars and amino acids

Safflower extract treatment elevated both reducing sugars (429.22 and 295.24 mg $100g^{-1}$ f.wt.) and amino acids (87.78 and 99.74 mg $100q^{-1}$ f.wt.) in chamomile shoots during the two seasons, respectively (Tables 6 and 7). Application with other plant extracts clearly induced an increase in reducing sugars and amino acids in comparing to plants sprayed with water.

Table 6. Effect of foliar application of plant aqueous extracts on some biochemical constituents in chamomile shoot during the season of 2016/ 2017

Data were presented as mean ± SD (n=3). Means in the same row with different letters are significantly different at P≤0.05

Table 7. Effect of foliar application of plant aqueous extracts on some biochemical constituents in chamomile shoot during the season of 2017/ 2018

Second season 2017 / 2018						
Biochemical	Control	Roselle	Turmeric	Safflower	Red beet	
constituents		extract	extract	extract	extract	
Chl a $(mg g^{-1} f.wt.)$	0.55° ±0.01	0.60° ±0.001	0.65° ±0.003	0.85° ±0.85	$0.71^b \pm 0.002$	
Chl b $(mg g^{-1} f.wt.)$	$0.22^{\rm e}$ ±0.002	0.25° ±0.003	0.25° ±0.001	$0.29^a \pm 0.002$	$0.28^b \pm 0.003$	
Carotenoids $(mg g^{-1} f.wt.)$	$0.20^{\rm e}$ ±0.001	0.21° ±0.004	0.22° ±0.003	$0.24^a \pm 0.004$	$0.23^b \pm 0.002$	
Flavonoids $(mg 100g^{-1}$ f.wt.)	15.32° ±0.03	89.63^{b} ±0.08	$101.20^a \pm 0.1$	22.40° ±0.05	39.66° ±0.02	
Phenolic compounds $(mg 100g^{-1}$ f.wt.)	$85.96^{\circ}+0.025$	89.12 ^b ±0.025	$103.76^a \pm 0.23$	55.76° ±0.02	64.33^{d} ±0.02	
RS (mg 100g ⁻¹ f.wt.)	198.36 ^e ±0.03	215.46° ±0.08	221.34° ±0.01	295.24 ^a ±0.04	253.63^{b} ±0.08	
AA (mg $100g^{-1}$ f.wt.)	65.48° ±1.22	71.67° ±1.27	77.39° ±1.73	$99.74^a \pm 0.03$	87.73^{b} ±1.12	

Data were presented as mean ± SD (n=3). Means in the same row with different letters are significantly different at P≤0.05

3.4 Effect of Plant Aqueous Extracts on Biochemical Constituents of Chamomile Inflorescence

Exogenous spray with red beet extract improved the total soluble phenolic compounds (601 and 563 mg 100 g⁻¹ d.wt.) and flavonoids (1282 and 773 mg 100 g $^{-1}$ d.wt.) in chamomile inflorescences followed by roselle or safflower extracts when compared with control, while spraying with turmeric extract led to reduce both phenols (483 and 420 mg 100 g^{-1} d.wt.) and flavonoids concentrations (485 and 296 mg 100 g^{-1} d.wt.) than control at both seasons, respectively (Tables 8 and 9).

Carotenoids concentration in chamomile inflorescences highly increased when safflower extract sprayed on plants which reached (40.64

and 37.48 mg 100 g^{-1} d.wt.) followed by red beet extract (36.78 and 34.01 mg 100 g⁻¹ d.wt.) which also showed a significant increase in carotenoids concentration more than control (28.94 and 27.55 mg 100 g^{-1} d.wt.) at both seasons, respectively.

3.5 Essential Oil Yield

Chamomile essential oil yield was increased when any of the tested plant aqueous extracts were applied in comparing to control (Tables 8 and 9). Essential oil yield increased to the maximum concentrations (2.43-2.25 and 2.07- 2.08 ml $100g^{-1}$ d.wt.) in inflorescences of chamomile sprayed with safflower and roselle extracts at the two seasons, respectively, in comparing with other extracts and water control as shown in Tables (8 and 9).

Table 8. Effect of foliar application of plant aqueous extracts on some biochemical constituents in chamomile inflorescences during the season of 2016/ 2017

First season 2016 / 2017						
Biochemical constituents	Control	Roselle	Turmeric	Safflower	Red beet	
		extract	extract	extract	extract	
Phenolic compounds $(mg 100g^{-1} d.wt.)$	$562^b \pm 10$	569° ±10	$483^{\overline{d}}\pm9$	538° ±1	$601^a \pm 5.29$	
Flavonoids $(mg 100g^{-1} d.wt.)$	574° ±4.36	712° ±10	$485^{\circ}+2$	942^{b} ±1	$1282^a \pm 9.5$	
Carotenoids		28.94° ±0.01 32.56 ^d ±0.01 33.45 ^c ±0.01		40.64° ±0.01	36.78^{b} ±0.01	
$(mg 100g^{-1} d.wt.)$						
Oil yield (ml $100g^{-1}$ d.wt.)	0.73° ±0.01	$2.07^b \pm 0.01$	1.19° ±0.01	2.43° ±0.01	1.22° ±0.01	
IC_{50} of essential oil (μ g ml ⁻¹)				14.36° ±0.03 35.82 ^d ±0.02 37.87 ^e ±0.026 20.74 ^c ±0.025	0.638^{a} ±0.002	
IC ₅₀ of inflorescences	1.465° +0.01		1.049° ±0.01 2.205 $^{\circ}$ ±0.001 0.349 $^{\circ}$ ±0.01		0.875° ±0.01	
ethanolic extract (mg ml^{-1})						

Data were presented as mean ± SD (n=3). Means in the same row with different letters are significantly different at P≤0.05

Table 9. Effect of foliar application of plant aqueous extracts on some biochemical constituents in chamomile inflorescences during the season of 2017/ 2018

Second season 2017 / 2018						
Biochemical constituents	Control	Roselle extract	Turmeric extract	Safflower extract	Red beet extract	
Phenolic compounds (mg $100g^{-1}$ d.wt.)	501° ±2	515° ±10	420° ±4.58	525° ±10	$563^a \pm 1$	
Flavonoids $(mg 100g^{-1} d.wt.)$	402° ±10	$592^b \pm 10$	296° ±3.6	$762^a + 10$	$773^a + 11$	
Carotenoids $(mg 100g^{-1} d.wt.)$	27.55° +0.01	31.32° ±0.01 31.98 ^c ±0.01		$37.48^a \text{±} 0.01$ $34.01^b \text{±} 0.01$		
Oil yield (ml 100g ⁻¹ d.wt.)	$0.58^{\rm e}$ ±0.01	$2.08^{\rm b}$ ±0.01	1.12° ±0.01	2.25° ±0.01	0.801° ±0.001	
IC ₅₀ of essential oil (µg ml ⁻¹) 20.61 ^b ±0.02 60.26 ^d ±0.06 88.83 ^e ±0.07					25.52° ±0.15 0.856 ^a ±0.004	
IC_{50} of inflorescences	1.345° ±0.016 0.939 $^{\circ}$ ±0.01		$1.512^{a} + 0.01$	0.235° ±0.01 0.778° ±0.01		
ethanolic extract (mg ml^{-1})						

Data were presented as mean ± SD (n=3). Means in the same row with different letters are significantly different at P≤0.05

3.6 Biochemical Constituents of Chamomile Essential Oil

GLC-MS analysis improved that chamomile essential oil contains 16 compounds *viz*, artimisia ketone, artimisia alcohol, iso-borneol, trans-β-farnesene, germacrene D, germacrene B, cadinene, sapthulenol, farnesene epoxide, tau-cadinol, α bisabolol oxide B, caryphylene oxide, bisbolone oxide, α bisabolol, chamazulene and α bisabolol oxide A (Tables 10 and 11). The main components of chamomile volatile and essential oil as detected by GC-MS were bisabolol oxide A, α bisabolol oxide B, trans – βfarnesene, chamazulene, bisabolone oxide organized by concentration for the two growing seasons.

 α Bisablol oxide A was the major compound that detected in chamomile volatile oil which increased with all exogenous spray treatments at the two growing seasons. The concentration of bisabolol oxide A reached about 90% in the essential oil produced from plants sprayed with safflower and red beet extracts, where safflower extract recorded the highest value (90.87%) followed by red beet extract 90.16%, roselle extract (89.25%), turmeric extract (80.74%) while control treatment gave the lowest value (78.64%) during the first season (Table 10), on the other hand, turmeric extract treatment gave the lowest value (78.88%) during the second season (Table 11).

At the first season, artimisia ketone, artimisia alcohol, trans-β-farnesene*,* bisabolone oxide and chamazulene were reduced when chamomile plants were sprayed with the tested plant extracts compared with control treatment, on the other hand, artimisia alcohol, artimisia ketone were increased in volatile oil when plants sprayed with turmeric or red beet extract on the second season. α Bisabolol was not detected with foliar spray of roselle, turmeric and safflower extracts at the first season, while it was detected with roselle and turmeric extract treatments at the second season. Cadinene, which not detected in untreated plants, was found in oil of chamomile treated with roselle (0.18 – 0.08%) and safflower extract $(0.22 - 0.12%)$ at both seasons, respectively. Tau-cadinol showed the highest value in safflower extract treatment (1.33% and 1.40%) while it showed the lowest value in red beet extract treatment (0.47% and 0.61%) during both seasons, respectively.

3.7 Scavenging Activity of Essential Oil and Ethanolic Extracts of Chamomile Inflorescences

Increasing the concentration of chamomile essential oil accompanied with increasing the scavenging activity on DPPH radical in all treatments (Figs. 1 and 2). Essential oil from plants treated with red beet extract had the highest scavenging activity on DPPH radical and

lowest value of IC₅₀ (0.638 and 0.856 μ g ml⁻¹) during 2017 and 2018, respectively, in comparison with other treatments (Fig. 1 and Tables 8 & 9).

Table 11. Effect of foliar application with plant aqueous extracts on the biochemical composition of essential oil in chamomile inflorescences and their percentages during the season of 2017/ 2018

Data is presented as percentage

Abdellatif and Ibrahim; AJSSPN, 4(1): 1-16, 2019; Article no.AJSSPN.46028

Fig. 2. Effect of foliar application of plant aqueous extracts on scavenging activity % of chamomile inflorescences ethanolic extract on DPPH radical during the season of 2017/2018

While ethanolic extract of chamomile inflorescences treated with safflower extract showed the highest radical scavenging activity and lowest IC_{50} (0.349 and 0.235 mg ml⁻¹) during 2016/2017 and 2017/2018, respectively, followed by red beet, roselle while turmeric aqueous extract treatment showed the lowest scavenging activity on DPPH radical and the highest IC_{50} value $(2.205$ and 1.512 mg ml⁻¹), respectively, as showed in Fig. 2 and Tables 8 & 9.

4. DISCUSSION

In this study, spraying plant aqueous extracts of roselle, turmeric, safflower and red beet on chamomile plants showed high efficiency in growth promotion and essential oil production. This growth stimulation may be due to the high content of sugars, amino acids and various secondary metabolites in these plant aqueous extracts. Red beet and safflower extracts contained the highest concentrations of soluble phenolic compounds, flavonoids, reducing sugars (RS), free amino acids (AA) in addition to

N, P, K % in the present study. These results were in agreement with Jasna et al. [32] who stated that red beet contained high concentrations of phenols, flavonoids, ß-cyanins and ß-xanthins beside the presence of sugars and protein that naturally exist in red beet. Also, Al Surmi et al. [33] recorded high concentrations of soluble phenols, amino acids and N, P, K ratios in safflower extract while roselle leaves and calyces contained phenols, flavonoids and anthocyanins that act as antioxidants [34]. Okereke et al. [35] reported that roselle extract contained a high value of glycosides. The main ingredients in roselle extract are vitamins C, A, D, B_1 and B_2 , antioxidants, anthocyanins, Fe, Mg and omega 3-ß-carotene [36].

Spraying apple trees with turmeric extract increased leaf nitrogen, phosphorus and potassium concentrations [37]. They attributed this increase to the high concentration of turmeric extract in potassium salt as found in this study. Ibrahim et al. [38] mentioned that both pH and titratable acidity were physical properties. The

value of pH and titratable acidity in the present study were compatible with that obtained by Ibrahim et al. [38] who found that roselle extract has a comparatively high acidic to cause lower pH in the extract. They attributed the high acidity of roselle to its natural constituents of organic acids such as citric acid, mallic acid and 3 indolyl acetic acid.

The physical properties and chemical constituents of the four studied aqueous extracts showed various economic traits in chamomile growth and productivity, roselle and turmeric extracts showed a clear increase in plant height while safflower and red beet extracts markedly provided large numbers of branches compared to other extract treatments. Similar findings were reported with alfa alfa, clover, red clover and landino aqueous extracts when influenced the growth of various legumes and grasses species [39]. Foliar spraying of pear tree "Le-Conte cv." with roselle, cinnamon and ginger water extracts gave the best fruits weight and fruits number per tree and increased total soluble solids and total fruit sugars % in comparison to control [40]. Based on the previous findings, it was cleared that using roselle extract improved the nutritional status, yield and physical-biochemical characteristics of Valencia orange fruits [41]. The higher content of K, vitamins, amino acids, curcumin and volatile oils in turmeric extract encourage researchers to interest in using it as an important plant extract [42]. The positive effect of turmeric extract on enhancing growth and productivity could be due to their higher content of protein, carbohydrates, amino acids, Ca, K, P, Fe, ascorbic acid, thiamine, riboflavin, niacin, curcumin and other pigments [8]. Also, Armanious [43] revealed that using turmeric extract at 0.05% was preferable than garlic and onion extracts in improving the leaf area, yield, nutrients status and fruit quality of Thompson seedless grapevines.

In the current study, the significant increase in growth and inflorescences production of plants sprayed with the aqueous extracts of safflower and red beet may be related by their high concentrations of reducing sugars and free amino acids which induced the highest values of growth parameters and gave higher yield components. These results were in agreement with Rolland et al. [44] who indicated that total sugars and amino acids serve as a storage sink involved in carbon and nitrogen pathways to modulate plant growth and development. Amino acids play various roles in plant physiological processes such as nitrogen source, hormone precursors, regulate nitrogen uptake that improved plant growth and yield [45]. Genetic analyses have approved extensive interaction between total sugars and plant hormones signaling activation [44]. Also, it was proved that accumulation of high levels of sugars in many plant species promote vegetative phase and increase number of leaves which resulted in elevating the canopy and the final outcome becomes increasing in the number of flowers, while low concentration of sugars slightly inhibited the flowering in arabidopsis [46].

Applied plant aqueous extracts as foliar spraying alleviated the concentrations of chl a, chl b and carotenoids in addition to increase reducing sugars and amino acids in treated chamomile plants in the present study. The explanation of this increase is that the active compounds in the chemical composition of the studied aqueous extracts especially in safflower and beet root extracts display potent antioxidant and osmoregulator properties under normal environmental conditions. In this context, Zonouri et al. [47] reported that increasing antioxidants in plant cells have a potential strength as free radical scavengers that prevent the degradation of chlorophylls and protect chloroplast membranes. Antioxidants can neutralize the $H₂O₂$ formation in the cell, which involved in abscisic acid (ABA) transmitting signals. ABA accelerated stomatal closure that limits the assimilation of $CO₂$ and affected photosynthesis process [48]. External application of antioxidants can reserve the stomatal closure [49]. Furthermore, Cushman [50] found that high levels of reducing sugars and amino acids induced an osmotic regulation in cells which improved the water absorbance and translocation that stabilize membranes and inhibit lipid peroxidation. These results also in the same trend with that obtained by Clifford et al. [51] who reported that the bioavailability of betalains and phenols, the main bioactive components in beet root, helping in protect cellular components in a state of redox balance under the normal metabolic conditions. Moreover, El Sharony et al. [52] stated that the main ingredients and antioxidant components of roselle extract have been shown to suppress oxide radical formation and increase total sugars, amino acids and ascorbic acid concentrations in mango fruits. The increasing in carotenoids in plants applied with aqueous extracts in the current study could be attributed to increase the biosynthesis of carotenoids and prevent the conversion of carotenoids into ABA under normal environmental conditions [48].

When biochemical analysis carried out in the inflorescences produced from plants sprayed with the tested plant extracts, it was observed that red beet extract encouraged the increase in soluble phenols, flavonoids and carotenoids in compared to other treatments that also provided these components more than control. Moreover, treatments of safflower and roselle extracts gave more yield of essential oil than red beet extract treatment. It was suggested that the reason of this increasing in yield was due to treatment with safflower or roselle extract stimulated the conversions of phenols and flavonoids, where are considered secondary products, into other secondary metabolites that have an essential role in the composition of volatiles and essential oil of chamomile inflorescences but these conversions were less in red beet extract treatment. This suggestion was compatible with Figueiredo et al. [53] who reported that the valuable volatile compounds and essential oils consist of multiple phenolic compounds mixed with alkaloids and terpenoids substances. According to these authors, the differences in aroma in the floral oil result from the changeable among the different compounds of phenols and terpenoids which give the oil distinctive characteristics. As well Zheljazkov et al. [54] showed similar results for the quantity and quality of the essential oil, where the treatments with aqueous extracts of absinthe worm wood, lavender and wild bergamot led to increase the oil yield and the essential oil components of Native spearmint.

In the current study, plant extract treatments altered the chemical constituents of chamomile essential oil by increasing or decreasing some
component percentage. Some treatments component percentage. Some treatments caused disappear in some components, this may be due to plant extract treatments influenced the essential biosynthesis and the conversion of compounds to others as mentioned before. These results were agreed with many investigators, McKay and Blumberg [55] stated that active principle components in chamomile essential oil are α bisabolol oxide A and B and chamazulene. The main constituents in the chamomile essential oil were chamazulene, α bisabolol, bisabolol- A and B oxides and ßfarnesene while the minor constituents were α and ß-caryophylene, caryophylene oxide and spathulenol [56]. German chamomile volatile oil has 5% chamazulene and 50% α bisabolol oxide

A [57]. Zheljazkov [58] reported that the sage brush and juniper water extracts increased the concentrations of ß-caryophyllene and trans- ß farnesene in spearmint essential oil relative to the water treatment. The author also reported that the spearmint essential oil content was more valuable when Juniper water extract was applied.

The concentration of antioxidants required to decrease initial DPPH radical concentrations by 50% (IC₅₀) is a measurement widely adopted for evaluating the antioxidant activity, where lower IC50 has higher antioxidant power. The present data revealed that chamomile essential oil and inflorescences ethanolic extract obtained from plants sprayed with red beet extract observed the highest free radical inhibitory activity on DPPH, according to their IC_{50} values, followed by safflower and roselle extracts in compared to plants treated with distilled water or turmeric extract. Similar results showed by Firat et al. [59] who found that chamomile essential oil contains a high level of free radical scavenging capacity through their higher DPPH inhibition and lower value of IC_{50} . They attributed this positive scavenging activity of chamomile to their higher content of α-bisabolol oxide A which exhibited the higher antioxidant potential with lower IC_{50} compared to ß-farnesene and α-bisabolol. This is fully consistent with the results obtained in this study where α-bisabolol oxide a found to be the main compound in the chamomile composition that increased to more than double the concentration in control plants and reached to 90% in the volatile oil of chamomile with safflower and red beet extract treatments. Agatonovic-Kustrin et al. [59] reported that chamomile flower heads and leaves had the most prominent antioxidant activities which α bisabolol and its oxide, apigenin and
chamazulene being the most effective chamazulene being the most effective antioxidants that prevent rancidity.

5. CONCLUSION

All plant aqueous extract treatments had a noticeable positive effect on plant growth, flowers production, oil yield and quality of chamomile. Safflower extract followed by red beet extract exhibited the best growth and shoot biochemical composition which improved the inflorescences yield, essential oil yield and valuable chemical constituents of essential oil.

So, plant extracts could be considered as alternative method to the chemical compounds, which achieve sustainability of organic farming.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Berry M. The chamomiles. Pharm. J. 1995; 254:191-193.
- 2. Srivastava JK, Shankar E, Gupta S. Chamomile: A herbal medicine of the past with bright future. Mol Med Report. 2010: 3(6):895–901.
- 3. Anonymous. How to grow kenaf for profit. Research Division Rep. MANR, Ibadan, Nigeria; 1970.
- 4. Schippers RR. African indigenous vegetables: An overview of the cultivated species. Chatham, UK. National Resources Institute/ACP-EU Technical Centre of Agricultural and Rural Cooperation. 2000;1-214.
- 5. Wong P, Yusof S, Ghazah HM, Cheman YE. Physico-chemical characteristics of roselle (*Hibiscus sabdariffa* L.), Nutr. and Food Sci. 2002;32:68-73.
- 6. Babalola SO, Babalola AO, Aworh OC. Compositional attributes of Roselle (*Hibiscus sabdariffa* L.). J. Food Technol. Afric. 2001;6:133-134.
- 7. Sivananda S. Home remedies. The Yoga Vedanta University, Sivananda Nagar, India. 1958;233-235.
- 8. Ashraf K, Sultan S. A comprehensive review on *Curcuma longa* Linn.: Phytochemical, pharmacological, and molecular study. International Journal of Green Pharmacy. 2017;(Suppl)11(4):671– 685.
- 9. Camas N, Cirak C, Esendal E. Seed yield, oil content and fatty acid composition of some safflower (*Carthamus tinctorius* L.) grown in Northern Turkey condition. J. Fac. of Agric. 2007;22:98-104.
- 10. Pedreño MA, Escribano J. Studying the oxidation and antiradical activity of betalain from beetroot. Journal of Biological Education. 2000;35:49–59.
- 11. Patkai G, Barta J, Varsanyi I. Decomposition of anticarcinogen factors of the beetroot during juice and nectar production. Cancer Letters. 1997;114: 105–106.
- 12. De Zwart FJ, Slow S, Payne RJ, Lever M, George PM, Gerrard JA, Chambers ST. Glycine betaine and glycine betaine analogues in common foods. Food Chemistry. 2003;83:197–204.
- 13. Atamanova A, Brezhneva TA, Slivkin AI, Nikolaevskii VA, Selemenev VF, Mironenko NV. Isolation of saponins from table beetroot and primary evaluation of their pharmacological activity. Pharmaceutical Chemistry Journal. 2005; 39(12):650–652.
- 14. Dias MG, Camoes MFGFC, Oliveira L. Carotenoids in traditional Portuguese fruits and vegetables. Food Chemistry. 2009; 113:808–815.
- 15. Vali L, Stefanovits-Banyai E, Szentmihalyi K, Febel H, Sardi E, Lugasi A, Kocsis I, Blazovics A. Liver-protecting effects of table beet (*Beta vulgaris* var. *Rubra*) during ischemia-reperfusion. Nutrition. 2007;23: 172–178.
- 16. Žitňanová I, Ranostajová S, Sobotová H, Demelová D, Pecháň I and Ďuračková Z. Antioxidative activity of selected fruits and vegetables. Biologia. 2006;61:279–284.
- 17. A.O.A.C. Official methods of analysis of the association of official analytical Chemists $17th$ ed. Published by the Association of Official Analytical Chemists. USA; 2000.
- 18. Ackerson RC. Osmoregulation in cotton in response to water stress, II- leaf carbohydrate state in relation to osmotic adjustment. Plant Physiol. 1981;67:489- 493.
- 19. Shahidi F, Naczk M. Methods of analysis and quantification of phenolic compounds. Food phenolic: Sources, chemistry, effects and applications. Technomic Publishing Company Inc. Lancaster, PA. 1995;287- 293.
- 20. Marinova D, Ribarova F, Atanassova M. Total phenolic and total flavonoids in Bulgarian fruits and vegetables. J. Univ. Chem. Tech. Metall: 2005;40(3):255-260.
- 21. Du CT, Francis FJ. Anthocyanins of roselle (*Hibiscus sabdoriffa* L). J. Food Sci. 1973; 38:810-812.
- 22. Miller GL. Use of dinitrosalicylic acid reagent for determination of reducing sugar. Anal. Chem. 1959;31:426.
- 23. Jayeraman J. Laboratory manual in biochemistry. Wiley Eastern Ltd. New Delhi, India. 1985;107.
- 24. Piper CS. Soil and plant analysis, Inter Sciences, New York. 1950;48-110.
- 25. Black CV, Evans DD, Ersminger LE, White KL, Clark FE. Methods of soil analysis. Amer. Soc. Agron. Inc. Bull. Medison, Wisconsin, USA. 1965;891-1400.
- 26. Wilde SA, Corey RB, Lyer IG, Voigt GK. Soil and plant analysis for tree culture. Oxford & IBH Publishing Co., New Delhi. 1985;1-218.
- 27. Costache MA, Campeanu G, Neata G. Studies concerning the extraction of chlorophyll and total carotenoids from vegetables. Romanian Biotechnological Letters. University of Bucharest. 2012; 17(5):7702-7708.
- 28. Guenther E. The essentials oils. D. van Nostrand Company Inc. New York. 1961;3.
- 29. Gulluce M, Sokmen M, Sahin F, Sokmen A, Adiguzel A, Ozer H. Biological activities of the essential oil and methanolic extract of *Micromeria fruticosa* L, Druce ssp *serpyllifolia (*Bieb) PH Davis plants from the eastern Anatolia region of Turkey. Journal of the Science of Food and Agriculture. 2004;84(7):735-741.
- 30. Snedecor GW, Cochran WG. Statistical analysis methods $7th$ ed. Iowa State Univ. Press Ames, USA. 1986;90.
- 31. Gomez KA, Gomez AA. Statistical procedures for agricultural research. 2nd ed. John Wiley and Sons, New York. 1984; 20-29,329-389.
- 32. Jasna M, Vić-Brunet ČAN, Savatović SS, Ćetković GS, Vulić JJ, Djilas SM, S. Markov SL, Cvetković DD. Antioxidant and antimicrobial activities of beet root pomace extracts. Czech J. Food Sci. 2011;29(6): 575–585.
- 33. Al Surmi NY, El Dengawy RAH, Khalifa AH. Chemical and nutritional aspects of some safflower seed varieties. J Food Process Technol. 2016;7:585. DOI: 10.4172/2157-7
- 34. Abdellatif YMR, Ibrahim MTS. Nonenzymatic antioxidants potential in enhancing *Hibiscus sabdariffa* L. tolerance to oxidative stress. International Journal of Botany; 2018.
- 35. Okereke CN, Iroka FC, Chukwuma MO. Phytochemical analysis and medicinal uses of *Hibiscus sabdariffa*. International Journal of Herbal Medicine. 2015;2(6):16- 19.
- 36. Bruneton J. Farmacogenosia. Zaragoza ed. Acriba. 2001;294-296.
- 37. Al-Hadethi MEA, Al-Hamdany MHSh, AL-Dulaimi AST. Role of garlic and turmeric extract in the leaves mineral contents of apple trees. IOSR Journal of Agriculture and Veterinary Science. 2016;9(10):2319- 2372.
- 38. Ibrahim N, Lee TS, Rozaini MZH. Potential application of roselle extract in functional food emulsions J. Teknol. dan Industri Pangan. 2013;24(1):22-26.
- 39. Grant EA, Sallans WG. Influence of plant extracts on germination and growth of eight forage species. Journal of Grass and Forage Science. 1964;19:191–197.
- 40. Abd-El-Latif FM, El-Gioushy SF, Ismail AF and Mohamed MS. The impact of biofertilization, antioxidants and potassium silicate on fruiting aspects and fruit quality of "Le-Conte" pear trees. Middle East Journal of Applied Sciences. 2017;7(2):1- 13.
- 41. Ahmed FF, Mansour AEM, Montasser MAA, Merwad MA, Mostafa EAM. Response of valencia orange trees to foliar application of roselle, turmeric and seaweed extracts. J. of Applied Sciences Research. 2013;9:960-964.
- 42. Peter KV. Informatices on turmeric and ginger India Spices. 1999;36(2-3):12-14.
- 43. Armanious MKU. The synergistic effect of spraying some plant extracts with some macro and micro nutrients of Thompson seedless grapevines. International Journal of Plant & Soil Science. 2014;3(10):1290- 1301.
- 44. Rolland F, Baena-Gonzalez E, Sheen J. Sugar sensing and signaling in plants: Conserved and Novel Mechanisms. Annual Review of Plant Biology. 2006; 57:675-709.
- 45. Kowalczyk K, Zielony T. Effect of amino plant and Asahi on yield and quality of lettuce grown on Rockwool. Conf. of Biostimulators in Modern Agriculture, Warsaw, Poland; 2008.
- 46. Ohto MA, Fischer RL, Goldberg RB, Nakamura K, Harada JJ. Control of seed mass by APETALA. Proc. Natl. Acad. Sci. USA. 2005;102(8):3123–3128.
- 47. Zonouri M, Javadi T, Ghaderi N, Saba MK. Effect of foliar spraying of ascorbic acid on chlorophyll a, chlorophyll b, total Chlorophyll, carotenoids, Hydrogen peroxide, leaf temperature and leaf relative water content under drought stress in grapes. Bull. Env. Pharmacol. Life Sci. 2014;3(Special Issue):178-184.
- 48. Taiz L, Zeiger E. Plant physiology. 5th Edition, Sinauer Associates Inc., Sunderland. 2010;782.
- 49. Chen Z, Gallie DR. The ascorbic acid redox state controls guard cell signaling and stomatal movement. Plant Cell,

American Society of Plant Biologists. 2004; 16:1143-1162.

- 50. Cushman JC. Osmoregulation in plants: Implications for agriculture integrative and comparative biology. 2001;41(4):758–769.
- 51. Clifford T, Howatson G, West DJ, Stevenson EJ. The potential benefits of red beetroot supplementation in health and disease. Nutients. 2015;7(4):2801–2822.
- 52. El-Sharony TF, El-Gioushy SF, Amin OA. Effect of foliar application with algae and plant extracts on growth, yield and fruit quality of fruitful mango trees cv. Fagri Kalan. J Horticulture. 2015;2(4):1-6.
- 53. Figueiredo AC, Barroso JG, Pedro LG, Scheffe JJC. Factors affecting secondary metabolite production in plants: Volatile components and essential oils Factors affecting volatile and essential oil production in plants. Flavour Fragr. J. 2008;23:213–226.
- 54. Zheljazkov VD, AstatkieT, Horgan T, Srogers M. Effect of plant hormones and

distillation water on mints. Hort. Science. 2010;45:1338–1340.

- 55. McKay DL, Blumberg JB. A review of the bioactivity and potential health benefits of chamomile tea (*Matricaria recutita* L.) Phytother. Res. 2006;20:519-530.
- 56. Costescu CI, Hadaruga NG, Rivis A, Hadaruga DI, Lupea AX, Parvu D. Antioxidant activity evaluation of some *Matricaria chamomilla* L. extracts. Journal of Agroalimentary Processes and Technologies. 2008;14:417-432.
- 57. Sharafzadeh S, Alizadeh O. German and roman chamomile. Journal of Applied Pharmaceutical Science. 2011;1(10):1-5.
- 58. Zheljazkov VD. Effect of foliar application of methyl jasmonate and extracts of juniper and sage brush on essential oil yield and composition of 'native' spearmint. Hort Science. 2013;48(4):462–465.
- 59. Firat Z, Demirci F, Demirci B. Antioxidant activity of chamomile essential oil and main components. Nat. Volatiles & Essent. Oils. 2018;5(1):11-16.

© 2019 Abdellatif and Ibrahim; This is an Open Access article distributed under the terms of the Creative Commons Attribution *License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.*

> *Peer-review history: The peer review history for this paper can be accessed here: http://www.sdiarticle3.com/review-history/46028*