

Current Journal of Applied Science and Technology

39(2): 47-56, 2020; Article no.CJAST.54429 ISSN: 2457-1024 (Past name: British Journal of Applied Science & Technology, Past ISSN: 2231-0843, NLM ID: 101664541)

Profiling Secondary Metabolites of Cocoa (Theobroma cacao L.) Endophytic Fungi Lasiodiplodia pseudotheobromae PAK-7 and Lasiodiplodia theobromae TN-R-3 and Their Antimicrobial Activities

M. Chaithra^{1,2}, S. Vanitha^{1*}, A. Ramanathan¹, V. Jegadeeshwari³, V. Rajesh⁴, V. Hegde⁵ and E. Apshara⁶

> ¹Department of Plant Pathology, TNAU, Coimbatore, Tamil Nadu, India. ²Department of Plant Pathology, CPCRI, RS, Vittal, Karnataka, India. ³Department of Spices and Plantation Crops, TNAU, India. ⁴Department of Plant Biotechnology, CPMB&B, TNAU, India. ⁵Division of Plant Protection, CPCRI, Kasaragod, Kerala, India. ⁶Department of Horticulture, CPCRI, RS, Vittal, Karnataka, India.

Authors' contributions

This work was carried out in collaboration among all authors. Authors MC, SV and VH designed the study. Author MC performed the statistical analysis, wrote the protocol and wrote the first draft of the manuscript. Authors VJ and EA provided samples for analysis. Authors AR and VR managed the literature searches. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/CJAST/2020/v39i230496 <u>Editor(s):</u> (1) Dr. Maduike Chiehiura Onwubiko Ezeibe, Professor, Department of Veterinary Medicine, Michael Okpara University of Agriculture, Umudike, Abia State, Nigeria. <u>Reviewers:</u> (1) M. E. Makgatho, University of Limpopo, South Africa. (2) Maria Antonietta Toscano, University of Catania, Italy. (3) Aba-Toumnou Lucie, University of Bangui, Central African Republic. Complete Peer review History: <u>http://www.sdiarticle4.com/review-history/54429</u>

> Received 01 January 2020 Accepted 19 February 2020 Published 26 February 2020

Original Research Article

ABSTRACT

Aims: To determine the chemical composition of secondary metabolite of cocoa endophytic fungi *L. pseudotheobromae* PAK-7, *L. theobromae* TN-R-3 and their anti-oomycete activities. **Statistical Design:** Multivariate analysis.

Place and Duration of Study: Department of Plant Pathology, TNAU, Coimbatore, Tamil Nadu from April 2018 to December 2019.

Methodology: Lipophilic extracellular secondary metabolites were extracted using ethyl acetate as a solvent and their chemical composition was detected by Gas Chromatography-Mass Spectrometry (GC-MS) and identified by NIST library and Pub Chem databases.

Results: Metabolic profiling of cocoa endophytic fungi *L. pseudotheobromae* PAK-7 and *L. theobromae* TN-R-3 showed the presence of eleven peaks representing nine compounds. The most abundant compound observed were Acetic acid, 3-methyl-6-oxo-9oxabicyclo[3.3.1]non-2-yl ester, 2H-Pyran-2-one, tetrahydro-4-hydroxy-6-pentyl-, Melezitose, Ethyl à-d-glucopyranoside collectively representing 58.01% area. In comparison to *L. pseudotheobromae* PAK-7 GC-MS analysis of *L. theobromae* TN-R-3 exhibited the presence of 29 peaks. The most abundant compounds were dl-Mevalonic acid lactone, Methyl 6-O-[1-methylpropyl]-á-d-galactopyranoside, 2H-Pyran-2-one, tetrahydro-4-hydroxy-6-pentyl-, Melezitose, Ethyl à-d-glucopyranoside, 1,6-Anhydro-á-d-talopyranose collectively representing 60.47% of the total area.

Conclusion: Chemical compositions and anti-oomycete activities of crude secondary metabolites of *L. pseudotheobromae* PAK-7, *L. theobromae* TN-R-3 differed entirely depending on the property and abundance of bioactive metabolites.

Keywords: Endophyte; cocoa; metabolic profiling.

1. INTRODUCTION

Microorganisms interact with many organisms in different ways. The interaction may be symbiotic or mutualistic or pathogenic depending on the nature of the host [1]. A large group of fungi are plant pathogens, and they live as endophytes, asymptomatically within the intracellular spaces of leaves, roots and stem tissues [2]. Endophytes are transmitted both vertically (systemic) and (nonsystemic). horizontally The interaction between host and endophytes varies, in vertical transmission, they exhibit mutualism and horizontally transmitted endophytes exhibit antagonism to the host [3]. The endophytic fungi become pathogenic upon senescence. The plants colonized with endophytic fungi are adapted to abiotic and biotic stresses [4]. The tolerance of plants against biotic stress attributed to the production of secondary metabolites by endophytic fungi [5]. The extracellular secondary metabolites produced by endopytic fungi are nonessential compounds that serve as ecological advantage under certain an environmental conditions [6] and possess antimicrobial activity [7]. L. theobromae and L. pseudotheobromae are cosmopolitan fungi having wide host range and exhibits pathogenic and saprophytic or endophytic nature [8,9, 10,11]. The metabolite produced by and L. L. pseudotheobromae F2 strain theobromae SPMKF 5 showed antibacterial activity [7,12]. Not much information is available on antimicrobial activity of metabolites produced by Lasiodiplodia sp. With this limited information, an attempt was made to characterize and screen

secondary metabolites produced by endophytic *L. pseudotheobromae.* PAK-7 and *L. theobromae* TN-R-3 for their anti-oomycete activity.

2. MATERIALS AND METHODS

2.1 Fungal Culture and Growth

In cocoa, black pod rot is a major problem and it is caused by Phytophthora palmivora. The virulent isolate of P. palmivora AYLB3 was selected for this study. Endophytic fungal isolates, isolated from cocoa were screened for their antagonistic activity against P. palmivora (MN814835- unpublished data) under in vitro conditions. Among all tested isolates. L. pseudotheobromae PAK-7 (MN418007) and L. theobromae TN-R-3 (MN400974) showed a cent percent inhibition (unpublished result). To check whether they are having any effect on cell wall lysis, the fungal cultures were grown in a 250 ml conical flask containing 100ml of malt extract broth and incubated at 27±2°C for 7 days inside a BOD incubator.

2.2 Secondary Metabolite Extraction

The ethyl acetate method of secondary metabolite extraction procedure was followed [13]. Endophytic fungal culture was grown in ME broth for seven days. The culture filtrate was collected by centrifuging at 6000 rpm for 20 min. An equal volume of culture filtrate and ethyl acetate was taken in a separating funnel and agitated for 10 min. The samples were extracted two to three times with ethyl acetate and evaporated. A brown colored concentrate secondary metabolite found after evaporation was dissolved in 1ml HPLC grade methanol.

2.3 Agar Well Diffusion Assay

The anti-oomycete activity of crude secondary metabolite of L. pseudotheobromae PAK-7 and L. theobromae TN-R-3 against Phytophthora palmivora AYLB3(MN814835- unpublished data) was evaluated by performing agar well diffusion method. Agar wells of 5 mm size were made using sterile cork-borer. Different concentration of secondary metabolites viz., 10, 20, 30, 40 µl was poured into the wells and sterile water was used as control. The experiment was repeated three times and inoculated plates were kept at 28°C for 48-72h [14]. The anti-oomycete activity was confirmed by the formation of a clear zone around the well. The experiment was repeated two times to confirm the cell wall lysis properties of crude metabolites of endophytic Lasiodiplodia isolates.

2.4 Gas Chromatography-Mass Spectrophotometry

The crude metabolite extracted from endophytic fungi was subjected to GC-MS analysis to identify the different bioactive compounds present in it. The Clarus SQ 8C Gas Chromatography-Mass Spectrometer from Perkin Elmer, was engaged for analysis. The DB-5 MS capillary standard non - polar column with 0.25 mm dimension (Agilent Co., USA) with Helium as carrier gas (1 ml/min) was used. The bioactive compounds present in crude metabolite extract were identified using NIST libraries MS search version 2.2. The spectrum of the known components was compared with the spectrum of the known components stored in the inbuilt library.

2.5 Statistical Analysis

Metabolic profile of *L. pseudotheobromae* PAK-7 and *L. theobromae* TN-R-3 isolates were analyzed using JMP[®] software. Under the multivariate method, principal component analysis (PCA) was performed following an independent linear combination of factors. Identified PCs were expressed in a heat map using an online heat mapper website following the average linkage clustering method and spearman rank correlation distance measurement method.

3. RESULTS AND DISCUSSION

3.1 Agar Well Diffusion Assay

Crude secondary metabolites from endophytic fungal isolates L. pseudotheobromae PAK-7 and L.theobromae TN-R-3 were screened for their anti-oomycete activity following agar well diffusion method as described by Sharma et al. [15] (Fig. 1). The L. theobromae TN-R-3 extracellular secondary metabolite exhibited mycelia cell wall lysis at 30 µl and 40 µl concentrations whereas L. pseudotheobromae PAK-7 exhibited maximum mycelial cell wall lysis at 40 µl concentration. The anti-oomycete activity secondary metabolites of endophytic by Lasiodiplodia is not studied well. But few reports are there on antibacterial properties of secondary metabolites of Lasiodiplodia sp. Taufig and Darah [14] reported that crude metabolite of L. pseudotheobromae IBRL OS-64 exhibited antibacterial activity against 13 human pathogenic bacteria. Among them. MRSA ATCC 33591, Staphylococcus aureus and Streptococcus mutans a gram-positive bacteria found most susceptible with an inhibition zone of ≥21 mm. Chagas et al. [16] reported antimicrobial activity of L. theobromae isolates FHS074, FHS075 and FHS076. These three isolates found effective against Escherichia coli, pneumonia, Klebsiella Pseudomonas aeruginosa. Orlandelli et al. [17] employed a cupplate method to screen secondary metabolite of L. theobromae an endophyte of the medicinal plant Piper hispidum Sw. against human pathogen Enterococcus hirae. Е. coli, Micrococcus luteus, Salmonella typhi and Staphylococcus aureus. The crude metabolite of L. theobromae at 61.4 mg/ml concentration inhibited all tested human pathogens.

3.2 Metabolite Profiling of *L.* pseudotheobromae PAK-7 and *L.* theobromae TN-R-3

Crude metabolites extracted from endophytic fungal isolates were subjected to GC-MS analysis. On a total of 40, bioactive metabolites were detected and they were matched with NIST library compounds (Table 1, Fig. 2). The relative abundance of metabolites at different retention time (RT) was represented as peaks. Chemical analysis of *L. pseudotheobromae* PAK-7 revealed the presence of eleven peaks corresponds to the presence of nine compounds. The most abundant compound observed were Acetic acid, 3-methyl-6-oxo-9oxabicyclo[3.3.1]non-2-yl ester, 2H-Pyran-2one, tetrahydro-4-hydroxy-6-pentyl-, Melezitose, à-d-glucopyranoside Ethyl collectively representing 58.01% area. In comparison to L. pseudotheobromae PAK-7 GC-MS analysis of L. theobromae TN-R-3 exhibited the presence of 29 peaks. The most abundant compounds were dl-Mevalonic acid lactone, Methvl 6-O-[1-methylpropyl]-á-d-galactopyranoside, 2H-Pvran-2-one. tetrahvdro-4-hydroxy-6-pentyl-, à-d-glucopyranoside Melezitose. Ethyl collectively representing 60.41% of the total area.

These two cocoa endophytic isolates belonged to the same genus, but their metabolic profiling is entirely different. Compounds like dl-Mevalonic acid lactone, Methyl 6-O-[1-methylpropyl]-á-dgalactopyranoside, 2H-Pyran-2-one, tetrahydro-4-hydroxy-6-pentyl-, Melezitose, Ethyl à-dglucopyranoside and Octadecanoic acid are common among them. Qian et al. [10] reported that Lasiodiplodia sp. ME4-2 endophytic fungi of Viscum coloratum produced five aromatic compounds which included cvclo-(Trp-Ala). indole-3-carboxvlic acid (ICA). indole-3carbaldehyde, mellein and 2-phenylethanol. This finding is in agreement with the current finding where isolate L.theobromae isolate TN-R-3 produced Cyclohexanecarboxylic acid. 2hydroxy-, ethyl ester.

dl-Mevalonic acid lactone is a δ -lactone form of mevalonic acid, a precursor in the mevalonate pathway. Metabolites of the mevalonate (MVA) pathway are involved in signaling pathways that transduce endosymbiotic microbial signals in the Medicago truncatula [18]. In the present study both isolates i.e., L. pseudotheobromae PAK-7 and L. theobromae TN-R-3 produced dl-Mevalonic acid lactone. This compound might have played a role in endophytic colonization. Antifungal activity of (R)-5,6-dihydro-6-pentyl-2H-pyran-2one against *Penicillium* species was reported by Parker et al. [19]. The methanolic extract of Solanum torvum contained Methyl 6-O-[1-methylpropyl]-á-d-galactopyranoside, that showed cytotoxic effect on cancer cell lines [20].

Melezitose is a non-reducing tri-saccharide also known as melicitose. It is a common metabolite produced by both two isolates of Lasiodiplodia sp. This compound produced from the stem and leaves of Alhagi maurorum Medik (Camel thorn) reported to be used in ancient ayurvedic medicine for the treatment of anorexia, dermatosis, and leprosy [21]. Another compound common among L. pseudotheobromae PAK-7 and L. theobromae TN-R-3 was Ethyl à-dglucopyranoside. The synonyms for this compound is Ethyl hexopyranoside. Mahdi et al. [22] reported that fungus growing termite Macrotermes bellicosus is used in traditional medicine in Benin for the treatment of diarrhoea. pulmonary infection and infectious and inflammatory diseases and it's effective against pathogenic microbes. NMR spectra of a chemical compound from Macrotermes bellicosus revealed the presence of compounds viz.. 2-methylhydroquinone, benzohydroguinone, niacinamide. ethyl-hexopyranoside, 3.4dihydroxyphenethyl alvcol and N-[2-(3,4-Dihydroxyphenyl)ethyl]acetamide.



Lasiodiplodia pseudotheobromae PAK-7



Lasiodiplodia theobromae TN-R-3

Fig. 1. In vitro screening of crude secondary metabolites employing agar well diffusion method Note: Zone of clearance indicates cell wall lysis of Phytophthora palmivora AYLB3

RT	Compound		%Area	
		PAK-7	TN-R-3	
3.033	1-Nitro-2-acetamido-1,2-dideoxy-d-mannitol	0.285	0.000	
3.249	2,4-Dihydroxy-2,5-dimethyl-3(2H)-furan-3-one	0.212	0.000	
3.444	ñ-Tetrahydro-3-furanmethanol	2.045	0.000	
5.664	4H-Pyran-4-one, 2,3-dihydro-3,5-dihydroxy-6methyl	0.118	0.000	
6.245	2-Cyclohexen-1-one, 4,5-dimethyl-	1.330	0.000	
6.98	Fluroxypyr	0.660	0.000	
7.295	1.2-Benzenediol. 3-methyl-	3.228	0.000	
7.475	dl-Mevalonic acid lactone	0.389	7.470	
7.575	1.2.4-Cvclopentanetrione, 3-butvl-	1.123	0.000	
7.71	1.5-Dimethyl-6-oxa-bicyclo[3.1.0]hexane	1.510	0.000	
7 875	Benzenemethanol 4-hvdroxy-	0 140	0.000	
8.13	1.1-Cvclohexanedimethanol	0.390	0.000	
8 266	2-Nitrohent-2-en-1-ol	1 251	0.000	
8 406	Octan-2-one 3 6-dimethyl-	1 355	0.000	
8 546	1 4-Benzenediol 2-methyl-	2 061	0.000	
8 686	Oxirane 2-(chloromethyl)-2-cyclobutyl-	0.810	0.000	
8 766	3(2H)-Euranone dibydro-5-isopropyl	2 4 3 9	0.000	
8 941	Prenovdiazine	0.236	0.000	
0.256	Methyl 6-0-[1-methylpronyl]-á-d-galactonyranoside	0.200	14 773	
0.381	2-Chloro-5-methylbenzene-1 3-diol	3 4 5 9	0.000	
9.501	3-Methoxymethoxy-2 3-dimethylundec-1-ene	0 374	0.000	
9.571	1.3-Cyclobeyanediol 2.5-dimethyl-2-nitro-	1 501	0.000	
9.090	Formamide N-[1-[(1-cyano-2methylpropyl)bydroxyamino]butyl	1.001	0.000	
9.700	A cetic acid 3 methyl 6 oxo 0oxahicyclo[3 3 1]pop 2 vl ester	26 / 97	0.000	
10.101	Sucroso	20.407 0.222	0.000	
10.301	2H Dyran 2 one tetrahydro 4 hydroxy 6 pentyl	6 234	10 706	
10.792	Melezitose	0.234	0.256	
12 240	Ethyl à d glucopyranasida	7 570	9.200	
13.540	n tert Butylestechol	1 367	0.000	
14 019	p-tert-Dutyicatechoi	0.522	0.000	
14.910	Banzaldahuda 2 athavu	1 221	0.000	
14.900	Totradoconcio acid	1.231	0.000	
10.914	n Hevedeenneis asid	0.203	0.000	
19.92		0.007	0.000	
23.007		0.230	23.307	
27.515	GIDDELETIC ACIO	0.201	0.000	
27.090	Dexadecialle, 1, 1-Dis(douecyloxy)-	0.210	0.000	
20.220	2-Hyuroxy-4-methoxy-7-methyl-7,0,9,10,11,12,13, 14-0000000	0.215	0.000	
20 660	17 Deptatriacentano	0.210	0.000	
20.000		0.319	0.000	
3.200	Isomanoi Denzeneethenel 4 hydroxy	0.000	3.200	
9.010	Denzeneeunanoi, 4-nyuroxy-	0.000	0.909	
10.33	Dibutul abtalata	0.000	0.304	
19.425	Dibutyi phinalate	0.000	0.393	
3.055	1-Nillo-2-acelaniluo-1,2-ulueoxy-u-manniloi	0.000	0.400	
3.209	∠,4-Diriyuroxy-∠,3-urineuryi-3(∠⊓)-iuran-3-one Enderital	0.000	2.390	
4.014	El yllillul Deminiscle	0.000	2.084	
4.104	Kopinii ole	0.000	1.795	
4.4/4	vietnanamine, N-methoxy-	0.000	1.536	
4.704	1,2,4-Cyclopentanetrione, 3-methyl-	0.000	2.925	
5.059	5-AMINOIEVUIINIC ACIO	0.000	0.776	
5.649	4n-ryian-4-one, 2,3-ainyaro-3,5-ainyaroxy-6metnyi	0.000	1.044	
6.065	o-Acetyl-L-serine	0.000	0.371	

Table 1. Metabolic profiling of endophytic fungi L. pseudotheobromae PAK-7 andL. theobromae isolate TN-R-3

Chaithra et al.; C.	JAST, 39(2): 47-56,	2020; Article no.	CJAST.54429
---------------------	---------------------	-------------------	-------------

RT	Compound	%Area	
		PAK-7	TN-R-3
6.855	5-Hydroxymethylfurfural	0.000	1.926
7.335	2H-Pyran-2-one, 5,6-dihydro-6-propyl-	0.000	1.120
7.77	Indole	0.000	0.871
7.995	Ethanone, 1-(2-hydroxy-5-methylphenyl)-	0.000	0.631
8.13	Cyclohexanecarboxylic acid, 2-hydroxy-, ethyl ester	0.000	0.930
8.556	á-D-Glucopyranose, 4-O-á-D-galactopyranosyl-	0.000	0.447
11.822	à-d-6,3-Furanose, methyl-á-d-glucohexodialdo-1,4furanoside	0.000	0.700
14.198	1,6-Anhydro-á-d-talopyranose	0.000	3.139
14.418	Isopropyl palmitate	0.000	1.165
18.795	Geraldol	0.000	0.456
19.72	n-Hexadecanoic acid	0.000	0.990
24.922	Pyrrolo[1,2-a]pyrazine-3-propanamide, 2,3,6,7,8, 8a-hexahydro- 1.4-dioxo	0.000	0.519



Fig. 2. GC-MS chromatogram showing different peaks representing % area (a) *L. pseudotheobromae* PAK-7 (b) *L. theobromae* TN-R-3



Fig. 3. PCA of extracellular secondary metabolites obtained by ethyl acetate fraction and identified by GC-MS in *L. pseudotheobromae*. PAK-7 and *L. theobromae* isolate TN-R-3

3.3 Principle Component Analysis

To identify compounds having antifungal activity in crude metabolite of endophytic *L. pseudotheobromae* PAK-7, and *L. theobromae* isolate TN-R-3 a multivariate analysis was carried out based on correlation [23]. The dataset consisted of a 2×40 matrix (Table 2). The row represented two isolates of *Lasiodiplodia* sp. and column was represented by compounds. The PCA was performed and the PCs having eigenvalue \geq 1 were considered. The score plot and loading plot for secondary metabolites of *L*. pseudotheobromae PAK-7, and L. theobromae TN-R-3 were shown in Fig. 3. The first two main PCs accounted for 100% variance; 56.60% variance was attributed to PC1 and it was positively correlated with Benzaldehyde,3ethoxy-, Melezitose, Acetic acid, 3-methyl-6-oxo-9oxabicyclo[3.3.1]non-2-yl ester and negatively correlated with Gibberellic acid. PC2 accounted for 43.40% of the total variable and it was positively correlated with Ethyl à-dglucopyranoside and Benzeneethanol, 4hydroxy- (Table 2, Figs. 3,4,5).

 Table 2. Unique principle components identified in L. pseudotheobromae PAK-7 and

 L. theobromae TN-R-3 isolates of cocoa

SI. no	Compound	Pak-7	TN-R-3	Property	Reference	
1	Ethyl à-d-glucopyranoside	+	+	Antibacterial	Mahdi et al. [22]	
2	Gibberellic acid	-	+	Growth	Waqas et al. [24]	
				promotion and		
				salinity		
				tolerance		
3	Melezitose	+	-	Antidematosis	Duke et al. [21]	
4	Acetic acid, 3-methyl-6-oxo-	+	-	Antibacterial	Wali and Abed [25]	
	9oxabicyclo[3.3.1]non-2-yl					
	ester					
5	Benzaldehyde, 3-ethoxy-	+	-	Antifungal	Ahluwalia et al. [26]	
6	Benzeneethanol, 4-hydroxy-	-	+	Antifungal	Ahluwalia et al. [26],	
					Manilal et al. [27]	
					но	
		0	5	OH	но но	
н	~	ANS.	XX	ne ne	оно	
п	° °	I'm T		HO'		
	° °	1 mil	\sim	- CH ₂	но	
н	HO'		7		о <i>~</i> -он	
	·o* ¥ *o.	I H	CO_H		PIO O	
	H, O	CH ₃	2		HOW OH	
					ÓН	
Ethyl E d gluconyronosido		Cibborallia aaid			Molozitoso	
		Gibber	enic aciu		WIEIEZILUSE	







Acetic acid, 3-methyl-6-oxo 9 oxabicyclo[3.3.1]non-2-yl ester

Benzaldehyde, 3-ethoxy-

Benzeneethanol, 4-hydroxy-





Fig. 5. Heat map for principle components identified in endophytic fungal isolates L. pseudotheobromae PAK-7 and L. theobromae isolate TN-R-3 (Clustering Method- Average linkage method, Distance Measurement Method-Spearman rank correlation)

4. CONCLUSION

In the present investigation secondary metabolic profiling and anti-oomycetes activity of crude extracellular metabolite of endophytic fungi L. pseudotheobromae PAK-7 and L. theobromae TN-R-3 was evaluated. Even though they are from the same genus their metabolic profiling differed. Agar well diffusion method and PCA showed that compounds present in crude metabolite of these two endophytic fungal isolates are having cell wall lysis property. There are reports on the production of signaling molecule viz., jasmonic acid by Lasiodiplodia sp. [28] but the antifungal/anti-oomycete activity of crude metabolites of Lasiodiplodia is not known or not studied. So an attempt was made to isolate and characterize secondary metabolites of endophytic fungal species of cocoa for their anti-oomycete activity. For further confirmation, investigations may be carried out to establish the anti-oomycete activity by the chemical compounds produced by these endophytic fungi.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Zilber-Rosenberg I, Rosenberg E. Role of microorganisms in the evolution of animals and plants: The hologenome theory of evolution. FEMS Microbiol Rev. 2008; 32(5):723-735.
- Wilson D. Endophyte The evolution of a term and clarification of its use and definition. Oikos.1995;73:274–276.
- Schardl CL, Liu JS, White JF, Finkel RA, An Z, Siegel MR. Molecular phylogenetic relationships of nonpathogenic grass mycosymbionts and clavicipitaceous plant pathogens. Plant Syst Evol. 1991;178(1): 27–41.
- Arnold AE, Mejía LC, Kyllo D, Rojas EI, Maynard Z, Robbins N, Herre EA. Fungal endophytes limit pathogen damage in a tropical tree. Proc Natl Acad Sci. 2003; 100(26):15649–15654.
- Saikkonen K, Faeth SH, Helander M, Sullivan TJ. Fungal endophytes: A continuum of interactions with host plants. Annu Rev Ecol Syst.1998;29(1):319–343.
- Leylaie S, Zafari D. Antiproliferative and antimicrobial activities of secondary metabolites and phylogenetic study of

endophytic *Trichoderma* species from Vinca plants. Front Microbiol. 2018;9:1484.

- Wei W, Jiang N, Mei YN, Chu YL, Ge HM, Song YC, Tan RX. An antibacterial metabolite from *Lasiodiplodia pseudotheobromae* F2. *Phytochemistry*. 2014;100:103-109.
- Kwon JH, Choi O, Kang B, Lee Y, Park J, Kang DW, Han I, Kim J. Identification of *Lasiodiplodia pseudotheobromae* causing mango dieback in Korea. Can J Plant Pathol. 2017;39:241–245.
- Alves A, Crous PW, Correia A, Phillips AJL. Morphological and molecular data reveal cryptic speciation in *Lasiodiplodia theobromae*. Fungal Diversity. 2008;28:1– 13.
- Qian C, Fu Y, Jiang F, Xu Z, Cheng D, Ding B, Gao C, Ding Z. *Lasiodiplodia* sp. ME4-2, an endophytic fungus from the floral parts of *Viscum coloratum*, produces indole-3-carboxylic acid and other aromatic metabolites. BMC Microbiol. 2014;14:297.
- 11. EI-Hawary SS, Sayed AM, Rateb ME, Bakeer W, AbouZid SF, Mohammed R. Secondary metabolites from fungal endophytes of *Solanum nigrum*. Nat Prod Res. 2017;31(21):2568-2571.
- Sibero MT, Igarashi Y, Radjasa OK, Sabdono A, Trianto A, Zilda D S, Wijaya YJ. Sponge-associated fungi from a mangrove habitat in Indonesia: Species composition, antimicrobial activity, enzyme screening and bioactive profiling. Int. J. Aquatic Res. 2019;11(2):173-186.
- 13. Devi NN, Prabakaran JJ. Bioactive metabolites from an endophytic fungus *Penicillium* sp. isolated from *Centella asiatica*. Curr Res Environ Appl Mycol. 2014;4(1):34-43.
- 14. Taufiq MMJ, Darah I. Fungal endophytes isolated from the leaves of a medicinal plant, *Ocimum sanctum* Linn and evaluation of their antimicrobial activities. Afr J Microbiol Res. 2018;12(26):616-622.
- Sharma D, Pramanik A, Agrawal PK. Evaluation of bioactive secondary metabolites from endophytic fungus *Pestalotiopsis neglecta* BAB-5510 isolated from leaves of *Cupressus torulosa* D. Don. 3 Biotech. 2016;6(2):210.
- 16. Chagas MBO. Prazeres Dos Santos I. Silva LC, Nascimento da et al. cultivable Antimicrobial activity of with endophytic fungi associated Hancornia speciosa Gomes Bark. Open Microbiol J. 2017;11:179-188.

- 17. Orlandelli RC, Alberto RN, Almeida TT, Azevedo JL, Pamphile JA. *In vitro* antibacterial activity of crude extracts produced by endophytic fungi isolated from *Piper hispidum* Sw. J. Appl. Pharm. Sci. 2012;2(10):137-141.
- Venkateshwaran M, Jayaraman D, Chabaud M, Genre A, Balloon AJ, Maeda J, Forshey K, Os D, Kwiecien NW, Coon JJ, Barker DG, Ané J. A role for the mevalonate pathway in early plant symbiotic signaling. PNAS. 2015;112(31): 9781-9786.
- 19. Parker SR, Cutler HG, Jacyno JM, Hill RA. Biological Activity of 6-Pentyl-2H-pyran-2one and Its Analogs. J Agric Food Chem. 1997;45(7):2774-2776.
- 20. Panigrahi S, Muthuraman MS, Natesan R, Pemiah B. Anticancer activity of ethanolic extract of *Solanum torvum* sw. Int J Pharm Pharm Sci. 2014;93-8.
- 21. Duke JA. Duke's handbook of medicinal plants of Latin America. CRC Press; 2008.
- 22. Mahdi DH, Hubert J, Schubert A, Vissiennon Z, Ahyi V, Nieber K, Vissiennon C. Chemical composition of and in vitro investigation the antibacterial activity of identified compounds from fungus-growing termites Macrotermes bellicosus. Planta Med. 2019;85(18):1388-1389.
- 23. Lia X, Maoa Z, Wua Y, Hoc H and Hea Y. Comprehensive volatile organic compounds profiling of *Bacillus* species with biocontrol properties by head space solid phase microextraction with gas chromatography-mass spectrometry. Biocontrol Sci Tech. 2015;25(2):132– 143.
- 24. Waqas M, Khan AL, Kamran M, Hamayun M, Kang SM, Kim YH, Lee IJ. Endophytic fungi produce gibberellins and indoleacetic acid and promotes host-plant growth during stress. Molecules. 2012;17(9): 10754-73.
- 25. Wali K, Abed MM. Antibacterial activity of acetic acid against different types of bacteria causes food spoilage. J Food Technol Pres. 201 9;3(11):1-4.
- Ahluwalia V, Kumar J, Rana VS, Singh R, Sati OP, Walia S. Synthesis and antimicrobial activity of esters of 3-ethoxy-4-hydroxybenzaldehyde oxime. Toxicol Envi Chem. 2017;99(1):1-9.
- 27. Manilal A, Tsalla T, Zerdo Z, Ameya G, Merdekios B, John SE. Evaluating the

antibacterial and anticandidal potency of mangrove, *Avicennia marina*. Asian Pac J Trop Dis. 2016;16(2):136-40.

28. Eng F, Haroth S, Feussner K, Meldau D, Rekhter D, Ischebeck T, Brodhun F, Feussner I. Optimized jasmonic acid production by *Lasiodiplodia theobromae* reveals formation of valuable plant secondary metabolites. PloS one. 2016; 11(12):e0167627.

Peer-review history: The peer review history for this paper can be accessed here: http://www.sdiarticle4.com/review-history/54429

^{© 2020} Chaithra et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.