



Phytochemical and Pharmacological Values of Two Major Constituents of *Asparagus* Species and their Nanoformulations: A Review

Yasmin Sobhy¹, Mohamed S. Mady^{1*}, Suzan A. Mina¹, Yasmin Abo-zeid^{2,3}

¹Department of Pharmacognosy, Faculty of Pharmacy, Helwan University, Ain Helwan, Cairo, Egypt. 11795. ²Department of Pharmaceutics and Industrial Pharmacy, Faculty of Pharmacy, Helwan University, Ain Helwan, Cairo, Egypt. 11795. ³Helwan Nanotechnology Centre, Helwan University, Helwan, Cairo, Egypt. 11792.

*Corresponding author: Mohamed S. Mady, Department of Pharmacognosy, Faculty of Pharmacy, Helwan University, Ain Helwan, Cairo, Egypt, 11795. Tel.: +2237454284
E-mail address: mohamedsaid_1985@yahoo.com

Submitted on: 30-05-2022; Revised on: 12-06-2022; Accepted on: 18-06-2022

To cite this article: Sobhy, Y.; Mady, M. S.; Mina, S. A.; Abo-zeid, Y. Phytochemical and Pharmacological Values of Two Major Constituents of *Asparagus* Species and their Nanoformulations: A Review. *J. Adv. Pharm. Res.* **2022**, 6 (3), 94-106. DOI: [10.21608/aprh.2022.141715.1176](https://doi.org/10.21608/aprh.2022.141715.1176)

ABSTRACT

Objectives: This review article highlights two of the major chemical classes and their derivatives frequently isolated from different species of genus *Asparagus* and the diversity of their biological activities in addition to different applications of genus *Asparagus* in nanoformulation. The species belonging to this genus are well known for their nutritional and medicinal benefits and are considered one of the promising sources of biologically active natural compounds. Among the major constituent detected in the genus *Asparagus* are saponins and flavonoids to which most of the biological activities are attributed. **Methods:** This review includes a collection of articles between 2000 to 2022, reviewed by internationally accepted databases and scientific journals. **Results:** This review demonstrates the structural and biological diversities of fifty-three saponin aglycones and glycosides and nineteen flavonoids and flavonoid glycosides isolated from different *Asparagus* species highlighting their structural diversity along with the biological activities of the reviewed species highlighting their structural diversity along with the biological activities of the reviewed species. Moreover, the application of *Asparagus* extracts for green synthesis of metal-based nanoparticles to minimize the hazardous effect on the environment and humans observed with metal-based nanoparticles chemically synthesized. **Conclusion:** The structural and biological diversities and potency of the reviewed saponins and flavonoids isolated from *Asparagus* along with their use in the production of stable nanoformulations made them a perfect candidate for future drug discovery of new pharmaceutically active agents.

Keywords: *Asparagus*, Saponins, Polyphenols, Nanoformula

INTRODUCTION

Plants are considered the major source of powerful drugs that have been used either for medical treatment or used as a precursor for semi-synthesis of effective analogs against several diseases.

The Asparagaceae family is a large group of herbaceous plants including around 114 genera, and the most famous one is *Asparagus*, including other several genera, genus *Asparagus* is one of its well-known plant genera, it comprises about 300 species of herbaceous perennials and woody shrubs all over the world¹ such as *Asparagus*

INTRODUCTION

Plants are considered the major source of powerful drugs that have been used either for medical treatment or used as a precursor for semi-synthesis of effective analogs against several diseases. The Asparagaceae family is a large group of herbaceous plants including around 114 genera, and the most famous one is *Asparagus*, including other several genera, genus *Asparagus* is one of its well-known plant genera, it comprises about 300 species of herbaceous perennials and woody shrubs all over the world¹ such as *Asparagus plumosus*, *A. asparagoides*, *A. officinalis*, *A. racemosus*, and *A. falcatus*². Based on the color difference, the genus *Asparagus* is classified into green, white, purple-green, purple-blue, and pink *Asparagus*³. *Asparagus* has been used as a food material for a long time for its nutritional benefits and also used for its medicinal properties⁴. The nutritive benefits of the genus *Asparagus* over other vegetables are due to the presence of higher content of proteins, fats, vitamins, and minerals³. *Asparagus* species are naturally located mainly in three continents, Asia, Africa, and Europe and many of them have established economic value as ornamental shrubs i.e. *A. plumosus* and *A. virgatus* or for their pharmacological importance i.e. *A. racemosus*, and *A. adscendens*⁵. Several studies were conducted to evaluate the phytochemical and pharmacological value of *Asparagus* species, with findings that highlighted their chemical diversity which justified their medicinal importance as well. *Asparagus* species have numerous biological properties, such as antioxidant^{6,7}, antihepatotoxic⁸, anti-inflammatory⁹, antibacterial¹⁰, and immunostimulant activities¹¹. Phytochemical studies of the genus *Asparagus* extensively highlighted the prominent content of steroidal saponins and phenolics³. Among the chemical profile of the genus *Asparagus*, steroidal saponins are the main group of phytochemicals isolated and identified¹². Moreover, green *Asparagus* is a rich source of phenolic compounds¹³ which justify the reported potent antioxidant and cytotoxic activities¹⁴. Nanotechnology has greatly impacted the field of pharmaceuticals and drug delivery. Nanomedicine is the branch of medicine that use particles sized from 1 to 1,000 nm for either therapeutic or diagnostic purposes¹⁵⁻¹⁸. Nanomedicine as a drug delivery system was used to target the drug to a specific site and consequently overcome drug accumulation at off-target tissues and consequently side effects associated with drug administration¹⁹⁻²³. This made nanomedicines to be able to overcome the limitations of conventional therapy²⁴ such as high frequency of drug administration²⁰ improve the delivery of a hydrophilic drug into cells²⁵ improve the bioavailability of poorly soluble drugs, control/sustain drug release^{26,27} and aid crossing the blood-brain barrier^{28,29}.

Nanoparticles (NPs) have been applied previously for diagnosis, prevention, and treatment of several diseases such as viral infections with promising results^{27,30-33}, and also, they demonstrated a good antibacterial activity against multidrug resistant bacteria,^{17,34-39} and for treatment of cancer, Alzheimer's, tuberculosis, wound healing repairing damaged tissue⁴⁰⁻⁴², and inflammation⁴³.

Literatures reported nano formulations of natural products were characterized by a remarkable improvement of the stability of the prepared formula (due to protection of active ingredients from physical and chemical degradation), solubility, bioavailability, and thus biological activity, as well as reduction of toxicity^{15,44}. Metal-based nanoparticles such as gold and silver were extensively studied due to their unique physicochemical properties rendering them massively applied in different disciplines including chemistry, biology, and biochemical¹⁷. Silver nanoparticles are considered one of the most formulated nanoparticles for various biomedical applications⁴⁵. However, they were commonly synthesized by a chemical method, which includes the administration of toxic chemicals, high energy, and pressure for successful production of nanomaterials. Toxic materials are hazardous to the surrounding environment and Humans⁴⁶. Thus, researchers adopted other strategies for green synthesis of metal-based nanoparticles including silver nanoparticles to make benefit of unique properties of nanomaterials as well as minimize any harmful or toxic effects on the environment and humans¹⁷. Several researchers have reported the use of plant extracts for the green synthesis of nanoparticles⁴⁷ and among its best strategies are either nano-suspension or nano-emulsion of plant extracts^{48,49}.

This review study covering a period of 22 years, aims to evaluate the content of different species of the genus *Asparagus* of saponins and phenolic compounds and their pharmacological effect along with the reported nanoformulations prepared with their extracts. The findings of this review study are categorized into different subclasses of the studied compounds as well as the collection of the reported biological activities of *Asparagus* species and its nano-applications.

MATERIAL AND METHODS

Search criteria

Original articles, research papers published in journals and PubMed Central, Google scholars on *Asparagus* species, and medicinal uses were collected and studied. The recorded collected data were selected when the keywords "Phytochemical, pharmacological significance, nanoformulations, *Asparagus* species" were typed in the search engines.

RESULTS AND DISCUSSION

Several reports indicated that saponins especially those with steroidal nucleus, flavonoid, and phenols are among the major constituents isolated from the genus *Asparagus*. Several other minor classes such as alkaloids, tannins, minerals, and amino acids were also reported from *Asparagus* species⁵⁰⁻⁵².

Steroidal saponin isolated from genus *Asparagus*

The main saponin of *A. officinalis* L., and some other *Asparagus* species, is protodioscin (C₅₁H₈₄O₂₂), which is a derivative of diosgenin of the spirostane, and furostanoid type (Figure 1) in a glycoside form^{53, 54}. Structural activity relationships (SAR) of the steroidal saponins were extensively studied and the reports indicated that structural diversity of the saponin glycoside leads to functional alterations. The SAR of steroidal saponins were found to be due to the monosaccharide's residues constituting the sugar part

and their sequences, as well as to the structures of the aglycones.^{55, 56} According to previous reports, saponins identified from *Asparagus* species, have different substitutions in the sapogenin (Aglycone part) as well as differences in their types, linkage, and the number of sugar residues. This tremendous structural variability opens the gate for diverse bioactivities. Table 1-6 illustrates the steroidal saponin of the spirostane and the furostane type and their derivatives isolated from *Asparagus* species (Figure 1-2). We noticed that they are dominated by the hexacyclic spirostane nucleus. The chemical structure of this group of compounds has a perhydrocyclopentanophenanthrene nucleus. Most of the natural spirostane saponins have the R configuration at C-22. The spirostanes also differ in the configuration at C-25 and 27-Me which can acquire an R/S configuration⁵⁷. The pentacyclic furostanols are characterized by a hemiacetal at C-22 and a C-26 glycosidic bond. Other steroidal compounds of androstane and cholestane nuclei isolated from *Asparagus* species are illustrated in Table 7.

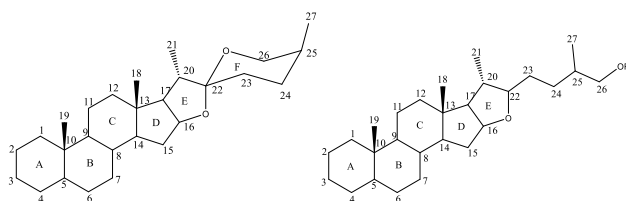


Figure 1. Spirostanol (left) and furostanol (right) saponins

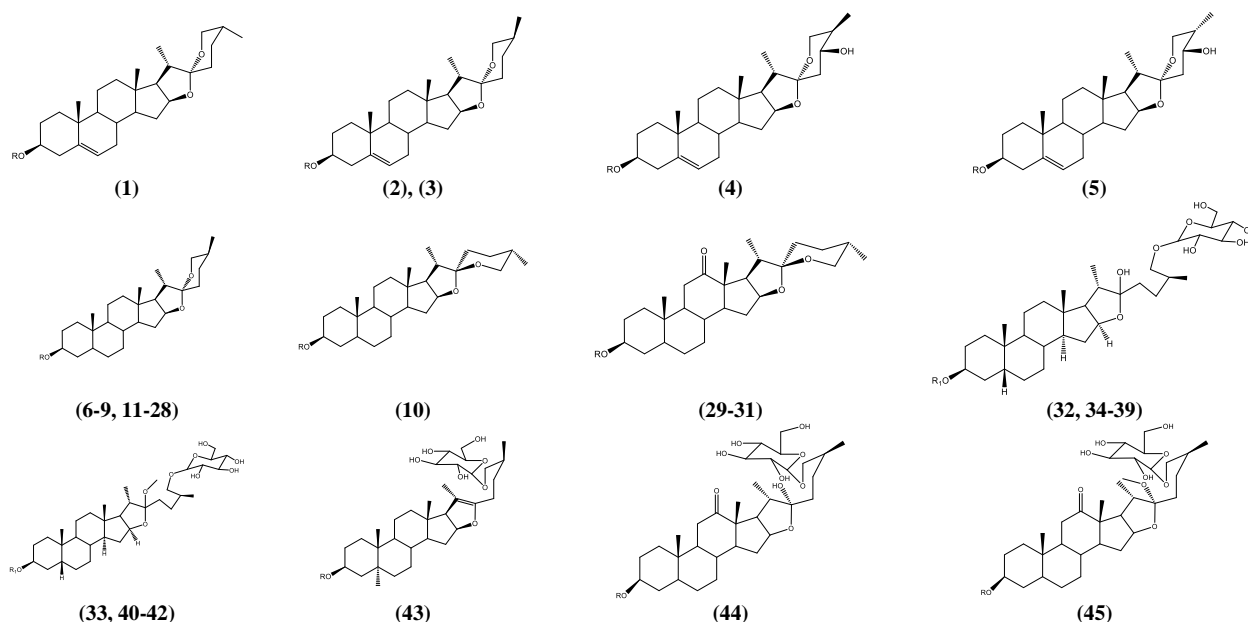


Figure 2. Spirostanoid, and furostanoid steroidal saponin isolated from *Asparagus* species.

Table 1. Spirosten saponin derivatives isolated from *Asparagus* species

Species(s)	Compound	R	Reference
<i>A. racemosus</i>	Diosgenin (25R) (1)	H	58
<i>A. Officinalis</i>	Yamogenin (25S) (2)		59
<i>A. adscendens</i>	Adscendin A (3)	β -D-Glu(1→6) α -L-Rha	60, 61
<i>A. cochinchinensis</i>	Asparagusoside C (4)	α -LRha-(1→2)-[α -L-Rha-(1→4)]- β -D-Glu	62
	Asparagusoside D (5)	α -L-Rha-(1→4)- β -D-Glu	

Table 2. Spirostan saponin derivatives isolated from *Asparagus* species

Species(s)	Compound	R	Reference
<i>A. cochinchinensis</i>	(25S)-5 β -spirostan-3 β -ol-3-O- β -D-glucopyranoside (6)	β -D-Glu	63
<i>A. racemosus</i>	Shatavarin IV	β -D-Glu(1→2) β -D-Glu (1→4) α -L-Rha	64
<i>A. adscendens</i>	(Asparanin B, Curillin H) (7)		
<i>A. racemosus</i>	Asparanin A (8)	β -D-Glu(1→2) β -D-Glu	1, 58, 65
	Shatavarin V (9)	β -D-Glu(1→2) α -L-Rha(1→4) β -D-Glu	
	Shatavarin VI (10)	β -D-Glu(1→2) β -D-Glu (1→4) α -L-Rha	
	Shatavarin VIII (11)	β -D-Glu(1→2) β -D-Glu(1→4) α -L-Ara(1→6) β -D-Glu	
	Shatavarin IX (12)	β -D-Glu(1→2) β -D-Glu(1→4) β -D-Glu	
	Shatavarin X (13)	β -D-Glu (1→2) α -L-Rha(1→4) β -D-6-acetylGlu	
	Shatavarside A (14)	β -D-Glu(1→2) α -L-Ara (1→6)Rha	1
	Shatavarside B (15)	β -D-Glu(1→2) β -D-Glu(1→4) α -L-Xyl(1→6) α -L-Rha	
	Immunoside (16)	β -D-Glu(1→2) α -L-Rha(1→4) α -L-Rha	1
<i>A. filicinus</i>	Filicinin A (17)	β -D-Glu(1→4)/ β -D-Glu(1→4) β -D-Gal(1→6)- β -D-Xyl	66
	Filicinin B (18)	β -D-Glu(1→4)/ β -D-Glu(1→2) β -D-Glu(1→4) β -D-Gal (1→6) β -D-Xyl	
<i>A. racemosus</i>	Filiasparoside C (19)	β -D-Glu(1→2) β -D-Xyl(1→4) α -L-Rha	1, 67, 68
<i>A. filicinus</i>	Filiasparoside D (20)	β -D-Glu (1→6) α -L-Ara	68
	Aspafilioside A (21)	β -D-Glu (1→4) β -D-Xyl	69, 70
	Aspafilioside B (22)	β -D-Glu(1→4) β -D-Xyl(1→6) α -L-Rha	
<i>A. racemosus</i>	Curillin H (23)	β -D-Glu (1→2) { α -L-Rha} (1→6)- β -D-Glu	71
<i>A. curillus</i>	Racemoside A (24)	β -D-Glu(1→4){ α -L-Rha (1→6)- β -D-Glu } (1→6) β -D-Glu	1, 72
	Racemoside B (25)	β -D-Glu (1→6){ α -L-Rha(1→6)- β -D-Glu }	
	Racemoside C (26)	β -D-Glu (1→4) α -L-Rha (1→6)- α -L-Rha	
<i>A. acutifolius</i>	(25S)-5 β - spirostan-3 β -ol-3-O- β -D-xylopyranosyl-(1→2)-[β -D-xylopyranosyl-(1→4)]- β -D-glucopyranoside (27)	β -D-Glu(1→2) β -D-Xyl (1→4)- β -D-Xyl	73
<i>A. africanus</i>	(25R)-5 β -spirostan-3 β -ol-3-O- β -D-glucopyranosyl-(1→2)-[α -L-arabinopyranosyl-(1→6)]- β -D-glucopyranoside } (28)	β -D-Glu(1→2) β -D-Glu(1→6)- α -L-Ara	74

Table 3. Spirostan-12-one saponin derivatives isolated from *Asparagus* species

Species(s)	Compound	R	Reference
<i>A. filicinus</i>	Filiasparoside A (29)	α -Glu(1→4) α -Xyl-(1→6)R-Ara	69,70
	Filiasparoside B (30)	α -Glu(1→6)R-Ara	
<i>A. africanus</i>	(25R)-3 β -hydroxy-5 β -spirostan-12-one 3-O- $\{\beta$ -D-glucopyranosyl-(1→2)- $[\alpha$ -L-arabinopyranosyl-(→6)]- β -D-glucopyranoside} (31)	β -D-Glu(1→2) β -D-Glu(1→6) α -L-Ara	74

Table 4. Saponins with furostan nucleus isolated from *Asparagus* species

Species(s)	Compound	R	Reference
<i>A. racemosus</i>	Shatavarin I (32)	β -D-Glu(1→2) β -D-Glu(1→6) α -L-Rha	1, 58
<i>A. curillus</i>	Curilloside G (33)	β -D-Glu(1→2) α -L-Rha (1→4)- β -D-Glu	71
<i>A. filicinus</i>	Aspafilioside C (34)	β -D-Glu (1→2)- β -D-Xyl (1→4) α -L-Ara	70
<i>A. acutifolius</i>	(25S)-3 β -5 β ,22 α -furostane-3,22,26-triol-3-O- β -D-xylopyranosyl(1→2)- $[\beta$ -D-xylopyranosyl(1→4)]- β -D-glucopyranosyl-26-O- β -D-glucopyranoside (35)	β -D-Glu (1→2)- β -D-Xyl (1→4) β -D-Xyl	73
<i>A. cochinchinensis</i>	(25S)-26-O- β -D-glucopyranosyl-5 β -furostan-3 β , 22 α ,26-triol-3-O- β -D-glucopyranoside (36)	β -D-Glu	63
	(25S)-26-O- β -D-glucopyranosyl-5 β -furostan-3 β , 22 α ,26-triol-3-O- α -L-rhamnopyranosyl-(1,4)- β -D-glucopyranoside (37)	β -D-Glu(1→4) α -L-Rha	
<i>A. curillus</i>	Curilloside H (38)	β -D-Glu(1→2) α -L-Rha (1→6) β -D-Glu	71
<i>A. racemosus</i>	Asparoside A (39)	β -D-Glu(1→2) β -D-Glu (1→4) α -L-Rha	
<i>A. africanus</i>	26-O- β -D-glucopyranosyl-22 α -methoxy-(25R)-furostan-3 β ,26-diol-3-O- $\{\beta$ -D-glucopyranosyl-(1→2)- β -D-glucopyranoside} (40)	β -D-Glu-(1→2) β -D-Glu	74
<i>A. acutifolius</i>	(25R)-3 β -5 β ,22 α -22-methoxyfurostane -3,26-diol-3-O- β -D-xylopyranosyl(1→2)- $[\beta$ -D-xylopyranosyl(1→4)]- β -D-glucopyranosyl-26-O- β -D-glucopyranoside (41)	β -D-Glu(1→2) β -D-Xyl (1→4) β -D-Xyl	73
<i>A. acutifolius</i>	(25R)-3 β -5 β ,22 α -22-methoxyfurostane -3,22 -diol-3-O- β -D-xylopyranosyl(1→2)- β -D-xylopyranosyl-26-O- β -D-glucopyranoside (42)	β -D-Glu(1→2) β -D-Xyl	73

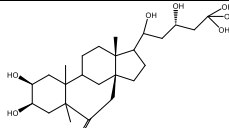
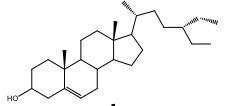
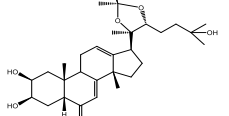
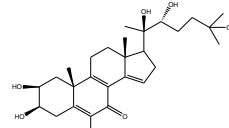
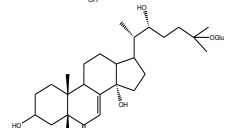
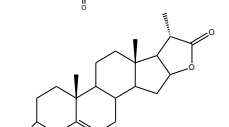
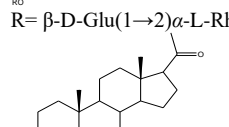
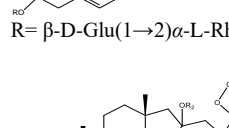
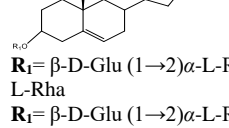
Table 5. Furostene saponin derivative isolated from *Asparagus* species

Species(s)	Compound	R	Reference
<i>A. filicinus</i>	Filiasparoside G (43)	β -D-Glu(1→4) β -D-Xyl(1→6) α -L-Ara	69

Table 6. Furostan-12-one saponin derivative isolated from *Asparagus* species

Species(s)	Compound	R	Reference
<i>A. cochinchinensis</i>	(25S)-26-O- β -D-glucopyranosyl-5 β -furostan-3 β ,22 α , 26-triol-12-one-3-O- β -D-glucopyranoside (44)	β -D-Glu	63
	(25S)-26-O- β -D-glucopyranosyl-22 α -methoxy-5 β -furostan-3 β , 26-diol-12-one-3-O- β -D-glucopyranoside (45)	β -D-Glu	

Table 7. Other steroidal compounds from *Asparagus* species

Species(s)	Compound	Structure	Reference
<i>A. filicinus</i>	Aspafilisine (46)		75
<i>A. officinalis</i>	β -Sitosterol (47)		59
<i>A. filicinus</i>	Ecdysterone [stachysterone- α -20,22-acetonide] (47)		60, 61
<i>A. dumosus</i>	Calonysterone (48)		76
	Blechnoside (49)		77
	Dumoside I (50)		
	Dumoside II (51)	 R = β -D-Glu(1 \rightarrow 2) α -L-Rha (1 \rightarrow 6) α -L-Rha	
	Dumoside III (52)	 R ₁ = β -D-Glu (1 \rightarrow 2) α -L-Rha (1 \rightarrow 6) α -L-Rha R ₂ = H	
	Dumoside IV (53)	 R ₁ = β -D-Glu (1 \rightarrow 2) α -L-Rha (1 \rightarrow 6) α -L-Rha R ₂ = CH ₃	

Flavonoids reported from different species of the genus *Asparagus*

Several reports have demonstrated scarce information regarding the genus *Asparagus* flavonoids and phenolic investigations. Fuentes *et al* 2007 investigated the phenolic content of both white and green varieties of the genus *Asparagus*, and reported that white *Asparagus* contained hydroxycinnamic acid derivatives, on the other hand, flavonoids were the major phenolics in green *Asparagus*⁷⁸. **Table 8** and **Figure 3** are illustrating different flavonoid classes identified from *Asparagus* species including classes such as

flavones (e.g., luteolin), flavanols (e.g., quercetin, kaempferol), flavanones (e.g., naringenin), and others.

Pharmacological studies

Several therapeutic potentials of *Asparagus* are well documented in the literature. The therapeutic activity is due to the possession of significant pharmacological activities such as antioxidant, anti-inflammatory, analgesic, antiulcer anti-aging, antifungal, and antimicrobial properties. **Table 9** describes the reported pharmacological activities of genus *Asparagus*.

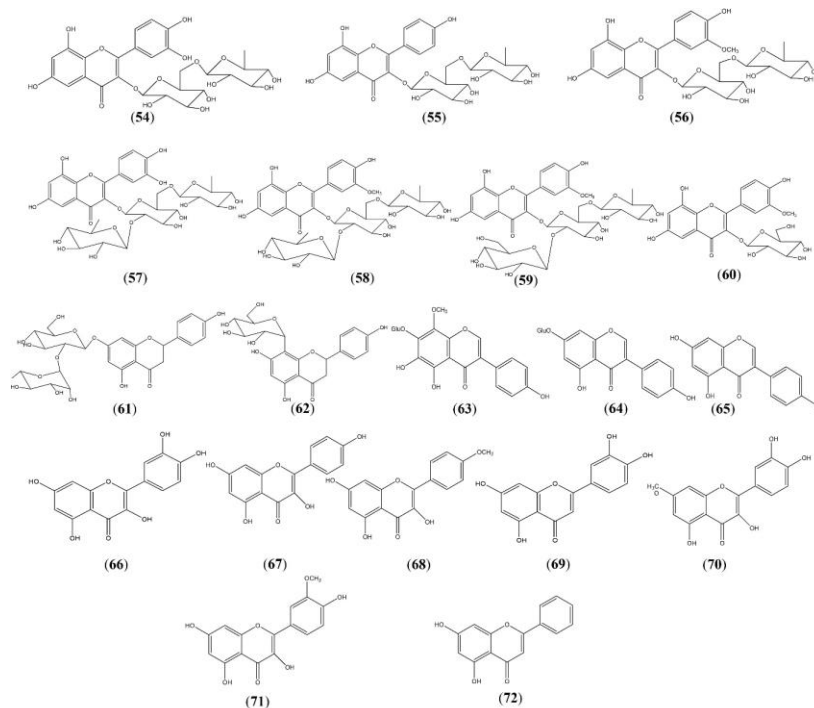


Figure 3. Flavonoids isolated from *Asparagus* species.

Table 8. Flavonoids reported from different *Asparagus* species

Compounds	Reference
Rutin (54)	1, 3, 78, 79
Kaempferol-3-O-rutinoside (55)	78
Isorhamnetin-3-O-rutinoside (56)	78
Quercetin-3-rhamnosyl-rutinoside (57)	79, 80
Isorhamnetin-3-rhamnosyl-rutinoside (58)	80
Isorhamnetin-3-glucosyl-rutinoside (59)	79, 80
Isorhamnetin-3-O-glucoside (60)	79, 80
Naringin (61), Vitexin (62)	81
8-Methoxy-5,6,4-trihydroxy isoflavone-7- O- β -glucopyranoside (63)	1
Genistin (64), Daidzein (65)	81
Quercetin (66)	1, 58, 78, 80
Kaempferol (67)	1, 78, 80, 81
Kaempferide (68), Luteolin (69)	81
Rhamnetin (70)	78
Isorhamnetin (71)	78, 79
Chrysin (72)	81

Nanoparticle formulation from extracts of genus *Asparagus*

Several reports have demonstrated the usage of the extracts of different species of the genus *Asparagus* in the synthesis of several metal-based nanoparticles due to the oxidative or reducing potential of active ingredients in the plant extracts¹⁰⁰. Moreover, the plant extract enriched with biologically active macromolecules (e.g. saponin, flavonoids, etc) was also encapsulated into nanoformulations such as liposomes, suspension, or alternatively adsorbed onto the surface of metal-based nanoparticles to act as a targeted drug delivery system to target the plant extract to diseased organ. Nanoformulations of plant extracts as previously discussed had superior advantages over conventional plant extract and it includes increasing the solubility, enhancing the bioavailability, reducing toxicity, and increasing the biological activity of the drug, in addition to increasing the sustainability and the protection against physical and chemical degradation¹⁰¹. However, literature reported that nanoparticles formulated using plant extracts for preparation are most likely to have biological activities similar to that of the original plant extracts but with the added benefit of optimizing the biological activity of the secondary metabolites¹⁰². Different nanoformulations prepared from extracts of different species of genus *Asparagus* and their biological activities were presented in **Table 10**.

Table 10. Biological activities of extracts of genus *Asparagus*

Biological activity	Species	Reference
Antioxidant	<i>A. racemosus</i>	82-84
	<i>A. cochinchinenss</i>	85
	<i>A. albus</i>	86
	<i>A. suaveolens</i>	87
	<i>A. stipularis</i>	88
Anti-aging	<i>A. cochinchinenss</i>	85
Anti-inflammatory	<i>A. racemosus</i>	58
	<i>A. pubescens</i>	89
	<i>A. cochinchinenss</i>	90, 91
	<i>A. africanus</i>	92
	<i>A. laricinus</i>	
Anti-microbial	<i>A. racemosus</i>	93-95
	<i>A. laricinus</i>	92
	<i>A. albus</i>	86
Anti-fungal	<i>A. retrofractus</i>	73, 96
	<i>A. acutifolius</i>	
	<i>A. setaceous</i>	
Anti-bacterial	<i>A. suaveolens</i>	87, 97
	<i>A. racemosus</i>	
Anti-ulcer	<i>A. pubescens</i>	98, 99
	<i>A. racemosus</i>	58, 98
Analgesic	<i>A. africanus</i>	92

Table 11. Nano formulations and biological significance of extracts of genus *Asparagus*

Species	Nanoformulation	Biological significance	Reference
<i>A. racemosus</i>	Silver nanoparticles	Antibacterial activity	103
		Bactericidal and Cytotoxic	104, 105
	Silver nanoparticles	Antibacterial and Immunomodulatory Potentials	100
	Gold nanoparticles		
	Silver nanoparticles	Ant mycobactericidal and Cytotoxicity	106
	Silver nanoparticles	Anti-diabetic activity	107
	Cobalt nanoparticles	Antibacterial activity	108
<i>A. officinalis</i>	Silver nanoparticles	Bactericidal and Cytotoxic	109
<i>A. adscendens</i>	Copper Nano-Particles	Antimicrobial Activities	110-112
<i>A. racemosus</i>	Liposomes	Anti-inflammatory activity	113
<i>A. stipularis</i>	Nanoencapsulation	Antioxidant	114

CONCLUSION

This review illustrated the structural diversity of two of the major classes of secondary metabolites separated from the genus *Asparagus*, including saponins and flavonoids. It also describes the diversity of their biological activities in addition to all nanoformulation application trials applied to different *Asparagus* extracts. The efficient comprehension of *Asparagus*'s steroidal saponins/flavonoids structural diversity and their biosynthetic pathways has scientific importance for further future studies of the possible use of the naturally occurring saponins and flavonoids as a chemical structural entity to prepare a library of semisynthetic derivatives for optimization of their biological activity and drug design research. Phytochemicals reported in *Asparagus* extracts were shown to be efficient in green synthesis of metal-based nanoparticles, where they can

reserve the biological activity of these nanoparticles but with no harmful effect on the environment and humans that was previously identified with metal-based nanoparticles synthesized chemically. In addition, nano-encapsulation of *Asparagus* extracts into one of the promising drug delivery systems e.g. liposomes, suspension, emulsion, etc was shown to have various merits compared to conventional *Asparagus* extracts such as improving solubility, stability, bioavailability, and hence, pharmacological activity.

Funding Acknowledgment

No external funding was received.

Conflict of interest

The authors declare that they have no conflicts of interest regarding the publication of this paper.

REFERENCES

1. Singla, R.; Jaitak, V., Shatavari (*Asparagus racemosus* wild): a review on its cultivation, morphology, phytochemistry and pharmacological importance. *Int. J. Pharm. Sci.* **2014**, *5*, 730-741.
2. Son, H. L.; Anh, N., Phytochemical composition, in vitro antioxidant and anticancer activities of quercetin from methanol extract of *Asparagus cochinchinensis* (LOUR.) Merr. tuber. *J. Med. Plant Res.* **2013**, *7*, 3360-3366.
3. Guo, Q.; Wang, N.; Liu, H.; Li, Z.; Lu, L.; Wang, C., The bioactive compounds and biological functions of *Asparagus officinalis* L. – A review. *J. Funct. Foods.* **2020**, *65*, 103727.
4. Benson, B. L., Update of the world's asparagus production areas, spear utilization and production periods. *Acta Hort.* **2002**, *589*, 33-40.
5. Stajner, N.; Bohanec, B.; Javornik, B., Genetic variability of economically important *Asparagus* species as revealed by genome size analysis and rDNA ITS polymorphisms. *Plant Sci.* **2002**, *162*, 931-937.
6. Yu, Q.; Fan, L., Antityrosinase and antioxidant activity of asparagus and its inhibition on B16F10 melanoma cells before and after hydrothermal treatment. *Food Bioscience.* **2021**, *41*, 101026.
7. Adouni, K.; Bendif, H.; Zouaoui, O.; Kraujalis, P.; Flamini, G.; Venskutonis, P. R.; Achour, L., Antioxidant activity of extracts obtained by high-pressure extraction procedures from *Asparagus stipularis* Forssk. *S. Afr. J. Bot.* **2022**, *146*, 789-793.
8. Acharya, S. R.; Acharya Ns Fau - Bhangale, J. O.; Bhangale Jo Fau - Shah, S. K.; Shah Sk Fau - Pandya, S. S.; Pandya, S. S., Antioxidant and hepatoprotective action of *Asparagus racemosus* Willd. root extracts. *Indian J Exp Biol.* **2012**, *50* (0019-5189 (Print)), 795-801.
9. Hassan, H.; Ahmadu, A.; Hassan, A., Analgesic And Anti-Inflammatory Activities Of *Asparagus africanus* Root Extract. *Afr J Tradit Complement Altern Med.* **2007**, *5*, 27-31.
10. Desoukey, S.; Nahas, S.; Sabh, A. Z.; Taha, Z.; El-Shabrawi, H., Antimicrobial effect of *Asparagus officinalis* l. extracts. *Plant Arch.* **2021**, *20*, 9253-9264.
11. Shanmugam, M.; H, G.; Lal, J.; S, C.; Kumar, D., Studies on the immunostimulant and antihepatotoxic activities of *Asparagus racemosus* root extract. *J. Med. Aromat. Plants.* **2001**, *22*, 49-52.
12. Onlom, C.; Nuengchamngong, N.; Phrompittayarat, W.; Putalun, W.; Waranuch, N.; Ingkaninan, K., Quantification of Saponins in *Asparagus racemosus* by HPLC-Q-TOF-MS/MS. *Nat Prod Commun.* **2017**, *12* (1), 7-10.
13. Karaaslan Ayhan, N.; Rosenberg, E., Development of comprehensive liquid chromatography with diode array and mass spectrometric detection for the characterization of (poly-)phenolic and flavonoid compounds and application to asparagus. *Food Chem.* **2021**, *354*, 129518.
14. Kobus-Cisowska, J.; Szymanowska-Powałowska, D.; Szczepaniak, O.; Gramza Michalowska, A.; Kmiecik, D.; Kulczyński, B.; Szulc, P.; Górnaś, P., Composition of polyphenols of asparagus spears (*Asparagus officinalis*) and their antioxidant potential. *Ciência Rural.* **2019**, *49*.
15. Ansari, S. H.; Islam, F.; Sameem, M., Influence of nanotechnology on herbal drugs: A Review. *J. Adv. Pharm. Technol. Res.* **2012**, *3* (3), 142-146.
16. Garnett, M. C.; Kallinteri, P., Nanomedicines and nanotoxicology: some physiological principles. *Occupational Med.* **2006**, *56* (5), 307-311.
17. Abo-zeid, Y.; Williams, G. R., The potential anti-infective applications of metal oxide nanoparticles: A systematic review. **2020**, *12* (2), e1592.
18. Abo-Zeid, Y.; Ismail, N. S. M.; McLean, G. R.; Hamdy, N. M., A molecular docking study repurposes FDA approved iron oxide nanoparticles to treat and control COVID-19 infection. *Eur J. Pharm. Sci: official J. Eur Federation Pharm. Sci.* **2020**, *153*, 105465-105465.
19. Moyle, G., Clinical manifestations and management of antiretroviral nucleoside analog-related mitochondrial toxicity. *Clinical Therapeutics.* **2000**, *22* (8), 911-936.
20. Abo-zeid, Y.; Williams, G. R.; Touabi, L.; McLean, G. R., An investigation of rhinovirus infection on cellular uptake of poly (glycerol-adipate) nanoparticles. *Int. J. Pharm.* **2020**, *589*, 119826.
21. Abo-zeid, Y.; Urbanowicz, R. A.; Thomson, B. J.; Irving, W. L.; Tarr, A. W.; Garnett, M. C., Enhanced nanoparticle uptake into virus infected cells: Could nanoparticles be useful in antiviral therapy? *Int. J. Pharm.* **2018**, *547* (1), 572-581.
22. Abo-zeid, Y.; Garnett, M. C., Polymer nanoparticle as a delivery system for ribavirin: Do nanoparticle avoid uptake by Red Blood Cells? *Journal of Drug Delivery Science and Technology.* **2020**, *56*, 101552.
23. Ritsema, J. A.; Weide, H. v. d.; Welscher, Y. M. t.; Goessens, W. H.; Nostrum, C. F. v.; Storm, G.; Bakker-Woudenberg, I. A.; Hays, J. P., Antibiotic-nanomedicines: facing the challenge of effective treatment of antibiotic-resistant respiratory tract infections. **2018**, *13* (15), 1683-1692.
24. Szunerits, S.; Barras, A.; Khanal, M.; Pagneux, Q.; Boukherroub, R., Nanostructures for the Inhibition of Viral Infections. *Molecules (Basel, Switzerland).* **2015**, *20* (8), 14051-14081.

25. Hillaireau, H.; Le Doan, T.; Appel, M.; Couvreur, P., Hybrid polymer nanocapsules enhance in vitro delivery of azidothymidine-triphosphate to macrophages. *J. Cont. Rel.* **2006**, *116* (3), 346-352.
26. Lembo, D.; Swaminathan, S.; Donalizio, M.; Civra, A.; Pastero, L.; Aquilano, D.; Vavia, P.; Trotta, F.; Cavalli, R., Encapsulation of Acyclovir in new carboxylated cyclodextrin-based nanosponges improves the agent's antiviral efficacy. *Int. J. Pharm.* **2013**, *443* (1), 262-272.
27. Burgess, K.; Li, H.; Abo-Zeid, Y.; Fatimah; Williams, G. R., The Effect of Molecular Properties on Active Ingredient Release from Electrospun Eudragit Fibers. *Pharmaceutics*. **2018**, *10* (3), 103.
28. Fiandra, L.; Colombo, M.; Mazzucchelli, S.; Truffi, M.; Santini, B.; Allevi, R.; Nebuloni, M.; Capetti, A.; Rizzardini, G.; Prosperi, D.; Corsi, F., Nanof ormulation of antiretroviral drugs enhances their penetration across the blood brain barrier in mice. *Nanomed. Nanotech Biol Med.* **2015**, *11* (6), 1387-1397.
29. Nowacek, A.; Gendelman, H. E., NanoART, neuroAIDS and CNS drug delivery. *Nanomedicine (London, England)*. **2009**, *4* (5), 557-574.
30. Henkel, M.; Geissler, M.; Weggenmann, F.; Hausmann, R., Production of microbial biosurfactants: Status quo of rhamnolipid and surfactin towards large-scale production. **2017**, *12* (7), 1600561.
31. Nii-Trebi, N. I., Emerging and Neglected Infectious Diseases: Insights, Advances, and Challenges. *BioMed Res. Int.* **2017**, *2017*, 5245021-5245021.
32. Garner, J. S.; Favero, M. S., Guideline for handwashing and hospital environmental control, 1985 supersedes guideline for hospital environmental control published in 1981. *Am. J. Infect. Cont.* **1986**, *14* (3), 110-126.
33. Pugliese, G., Recommendations for preventing the spread of vancomycin resistance. *Hosp. Epidemiol.* **1995**, *16* (0899-823X (Print)), 498.
34. Abo-zeid, Y.; Amer, A.; El-Houssieny, B.; Mahmoud, M.; Sakran, W., Overview on Bacterial Resistance and Nanoparticles to Overcome Bacterial Resistance %. *J Adv. Pharm. Res.* **2021**, *5* (3), 312
35. Wang, Y.; Zhang, Z.; Abo-zeid, Y.; Bear, J. C.; Davies, G.-L.; Lei, X.; Williams, G. R., SiO₂-coated layered gadolinium hydroxides for simultaneous drug delivery and magnetic resonance imaging. *J. Solid State Chem.* **2020**, *286*, 121291.
36. Chintagunta, A. D.; M, S. K.; Nalluru, S.; N S, S. K., Nanotechnology: an emerging approach to combat COVID-19. *Emergent Mater.* **2021**, 1-12.
37. Rangayasami, A.; Kannan, K.; Murugesan, S.; Radhika, D.; Sadasivuni, K. K.; Reddy, K. R.; Raghu, A. V., Influence of nanotechnology to combat against COVID-19 for global health emergency: A review. *Sensors Int.* **2021**, *2*, 100079-100079.
38. Abo-zeid, Y.; Bakkar, M. R.; Elkhoully, G. E.; Raya, N. R.; Zaafar, D., Rhamnolipid Nano-Micelles versus Alcohol-Based Hand Sanitizer: A Comparative Study for Antibacterial Activity against Hospital-Acquired Infections and Toxicity Concerns. **2022**, *11* (5), 605.
39. Bakkar, M. R.; Faraag, A. H. I.; Soliman, E. R. S.; Fouda, M. S.; Sarguos, A. M. M.; McLean, G. R.; Hebshy, A. M. S.; Elkhoully, G. E.; Raya, N. R.; Abo-zeid, Y., Rhamnolipids Nano-Micelles as a Potential Hand Sanitizer. **2021**, *10* (7), 751.
40. Hamdan, S.; Pastar, I.; Drakulich, S.; Dikici, E.; Tomic-Canic, M.; Deo, S.; Daunert, S., Nanotechnology-Driven Therapeutic Interventions in Wound Healing: Potential Uses and Applications. *ACS Cent. Sci.* **2017**, *3* (3), 163-175.
41. Meylina, L.; Muchtaridi, M.; Joni, I. M.; Mohammed, A. F. A.; Wathoni, N., Nanoformulations of α -Mangostin for Cancer Drug Delivery System. *Pharmaceutics*. **2021**, *13* (12), 1993.
42. Tocco, I.; Zavan, B.; Bassetto, F.; Vindigni, V., Nanotechnology-Based Therapies for Skin Wound Regeneration. *J. Nanomater.* **2012**, *2012*, 714134.
43. Abo-zeid, Y.; Diab, R.; Sanad, R.; Sakran, W., Recent Advances in Herbal-Based Nanomedicine for Anti-Inflammatory Purposes %J Journal of Advanced Pharmacy Research. **2021**, *5* (4), 387-397.
44. Bonifácio, B. V.; Silva, P. B. d.; Ramos, M. A. D. S.; Negri, K. M. S.; Bauab, T. M.; Chorilli, M., Nanotechnology-based drug delivery systems and herbal medicines: a review. *Int. J. Nanomed.* **2014**, *9*, 1-15.
45. Wijnhoven, S. W. P.; Peijnenburg, W. J. G. M.; Herberts, C. A.; Hagens, W. I.; Oomen, A. G.; Heugens, E. H. W.; Roszek, B.; Bisschops, J.; Gosens, I.; Van De Meent, D.; Dekkers, S.; De Jong, W. H.; van Zijverden, M.; Sips, A. J. A. M.; Geertsma, R. E., Nano-silver – a review of available data and knowledge gaps in human and environmental risk assessment. *Nanotoxicology*. **2009**, *3* (2), 109-138.
46. Gupta, P.; Mahajan, A., Green chemistry approaches as sustainable alternatives to conventional strategies in the pharmaceutical industry. *RSC Advances*. **2015**, *5* (34), 26686-26705.
47. Faisal, S.; Jan, H.; Shah, S. A.; Shah, S.; Khan, A.; Akbar, M. T.; Rizwan, M.; Jan, F.; Wajidullah; Akhtar, N.; Khattak, A.; Syed, S., Green Synthesis of Zinc Oxide (ZnO) Nanoparticles Using Aqueous Fruit Extracts of *Myristica fragrans*: Their Characterizations and Biological and Environmental Applications. *ACS Omega*. **2021**, *6* (14), 9709-9722.

48. El-Naggar, M. E.; Soliman, R. A.; Morsy, O. M.; Abdel-Aziz, M. S., Nanoemulsion of Capsicum fruit extract as an eco-friendly antimicrobial agent for production of medical bandages. *Biocatal. Agric. Biotechnol.* **2020**, *23*, 101516.
49. Gaur, P., Nanosuspension of flavonoid-rich fraction from *Psidium guajava* Linn for improved type 2-diabetes potential. *J Drug Deliv Sci Technol.* **2021**, *62*, 102358.
50. Olivier, M.; Muganza, F. M.; Shai, L. o. L.; Gololo, S.; Nmutavhanani, L., Phytochemical screening, antioxidant and antibacterial activities of ethanol extracts of *Asparagus suaveolens* aerial parts. *S. Afr. J. Bot.* **2017**, *108*, 41–46.
51. Hamdi, A.; Jaramillo-Carmona, S.; Rodríguez Arcos, R.; Jiménez, A.; Lachaal, M.; Karray-Bourouai, N.; Guillen, R., Phytochemical Characterization and Bioactivity of *Asparagus acutifolius*: A Focus on Antioxidant, Cytotoxic, Lipase Inhibitory and Antimicrobial Activities. *Molecules.* **2021**, *26*, 3328.
52. Hussain, A.; Ahmad, M. P.; Wahab, S.; Sarfaraj, M., A Review on Pharmacological and Phytochemical Profile of *Asparagus racemosus*. *Pharmacologyonline.* **2011**, 1353-1364.
53. Wang, M.; Tadmor Y Fau - Wu, Q.-L.; Wu Qi Fau - Chin, C.-K.; Chin Ck Fau - Garrison, S. A.; Garrison Sa Fau - Simon, J. E.; Simon, J. E., Quantification of protodioscin and rutin in asparagus shoots by LC/MS and HPLC methods. *J Agric Food Chem.* **2003**, *51(21)* (0021-8561 (Print)), 6132-6.
54. Lee, E. J.; Yoo, K. S.; Patil, B. S., Development of a Rapid HPLC-UV Method for Simultaneous Quantification of Protodioscin and Rutin in White and Green *Asparagus* Spears. *J Food Sci.* **2010**, *75* (9), C703-C709.
55. Mimaki, Y.; Yokosuka, A.; Kuroda, M.; Sashida, Y., Cytotoxic Activities and Structure-Cytotoxic Relationships of Steroidal Saponins. *Biol. Pharm. Bull.* **2001**, *24* (11), 1286-1289.
56. Hernández, J. C.; León, F.; Brouard, I.; Torres, F.; Rubio, S.; Quintana, J.; Estévez, F.; Bermejo, J., Synthesis of novel spirostane saponins and their cytotoxic activity. *Bioorg. Med. Chem.* **2008**, *16* (4), 2063-2076.
57. Van Minh, C.; Dat, N. T.; Dang, N. H.; Nam, N. H.; Ban, N. K.; Van Tuyen, N.; Huong, L. M.; Huong, T. T.; Van Kiem, P., Unusual 22S-spirostane Steroids from *Dracaena cambodiana*. *Nat. Prod. Commun.* **2009**, *4* (9), 1934578X0900400908.
58. Singh, R.; Geetanjali, *Asparagus racemosus*: a review on its phytochemical and therapeutic potential. *Nat. Prod. Res.* **2016**, *30* (17), 1896-1908.
59. Huang, X.; Kong, L., Steroidal saponins from roots of *Asparagus officinalis*. *Steroids.* **2006**, *71* (2), 171-176.
60. Negi, J. S.; Singh, P.; Joshi, G. P.; Rawat, M. S.; Bisht, V. K., Chemical constituents of *Asparagus*. *Pharmacogn. Rev.* **2010**, *4* (8), 215-220.
61. Sharma, S. C.; Sharma, H. C., Oligofuro- and spirostanosides of *Asparagus adscendens*. *Phytochem.* **1984**, *23* (3), 645-648.
62. Liu, B.; Li, B.; Zhou, D.; Wen, X.; Wang, Y.; Chen, G.; Li, N., Steroidal saponins with cytotoxic effects from the rhizomes of *Asparagus cochinchinensis*. *Bioorg. Chem.* **2021**, *115*, 105237.
63. Zhu, G.-L.; Hao, Q.; Li, R.-T.; Li, H.-Z., Steroidal saponins from the roots of *Asparagus cochinchinensis*. *Chinese J. Nat. Med.* **2014**, *12* (3), 213-217.
64. Singla, R.; Jaitak, V., SHATAVARI (ASPARAGUS RACEMOSUS WILD): A REVIEW ON ITS CULTIVATION, MORPHOLOGY, PHYTOCHEMISTRY AND PHARMACOLOGICAL IMPORTANCE. *Int. J. Pharm. Sci Res.* **2014**, *5*, 730-741.
65. Sharma, S. C.; Chand, R.; Sati, O. P., Steroidal saponins of *Asparagus adscendens*. *Phytochem.* **1982**, *21* (8), 2075-2078.
66. Sharma, S. C.; Thakur, N. K., Oligofurostanosides and oligospirostanosides from roots of *Asparagus filicinus*. *Phytochemistry.* **1996**, *41* (2), 599-603.
67. Sharma, U.; Saini, R.; Kumar, N.; Singh, B., Steroidal Saponins from *Asparagus racemosus*. *Chem. Pharm. Bull.* **2009**, *57* (8), 890-893.
68. Zhou, L.-B.; Chen, T.-H.; Bastow, K. F.; Shibano, M.; Lee, K.-H.; Chen, D.-F., Filiastanosides A–D, Cytotoxic Steroidal Saponins from the Roots of *Asparagus filicinus*. *J. Nat. Prod.* **2007**, *70* (8), 1263-1267.
69. Wu, J.-J.; Cheng, K.-W.; Zuo, X.-F.; Wang, M.-F.; Li, P.; Zhang, L.-Y.; Wang, H.; Ye, W.-C., Steroidal saponins and ecdysterone from *Asparagus filicinus* and their cytotoxic activities. *Steroids.* **2010**, *75* (10), 734-739.
70. Zhou, L.; Cheng, Z.; Chen, D., Simultaneous determination of six steroidal saponins and one ecdysone in *Asparagus filicinus* using high performance liquid chromatography coupled with evaporative light scattering detection. *Acta Pharmaceutica Sinica B.* **2012**, *2* (3), 267-273.
71. Hayes, P. Y.; Jahidin, A. H.; Lehmann, R.; Penman, K.; Kitching, W.; De Voss, J. J., Asparinins, asparosides, curillins, curillosides and shavatarins: structural clarification with the isolation of shatavarin V, a new steroidal saponin from the root of *Asparagus racemosus*. *Tetrahedron Lett.* **2006**, *47* (49), 8683-8687.
72. Mandal, D.; Banerjee, S.; Mondal, N. B.; Chakravarty, A. K.; Sahu, N. P., Steroidal saponins from the fruits of *Asparagus racemosus*. *Phytochem.*

- 2006, 67 (13), 1316-1321.
73. Sautour, M.; Miyamoto, T.; Lacaille-Dubois, M.-A., Steroidal saponins from *Asparagus acutifolius*. *Phytochem.* **2007**, 68 (20), 2554-2562.
74. Debella, A.; Haslinger, E.; Kunert, O.; Michl, G.; Abebe, D., Steroidal saponins from *Asparagus africanus*. *Phytochem.* **1999**, 51 (8), 1069-1075.
75. Wang, J.-P.; Cai, L.; Chen, F.-Y.; Li, Y.-Y.; Li, Y.-Y.; Luo, P.; Ding, Z.-T., A new steroid with unique rearranged seven-membered B ring isolated from roots of *Asparagus filicinus*. *Tetrahedron Lett.* **2017**, 58.
76. Khaliq-uz-Zaman, S. M.; Simin, K.; Ahmad, V. U., Chemical constituents from *Asparagus dumosus*. *Fitoterapia.* **2000**, 71 (3), 331-333.
77. Ahmad, V. U.; Khaliq-uz-Zaman, S. M.; Shameel, S.; Perveen, S.; Ali, Z., Steroidal saponins from *Asparagus dumosus*. *Phytochem.* **1999**, 50 (3), 481-484.
78. Fuentes-Alventosa, J. M.; Rodríguez, G.; Cermeño, P.; Jiménez, A.; Guillén, R.; Fernández-Bolaños, J.; Rodríguez-Arcos, R., Identification of Flavonoid Diglycosides in Several Genotypes of *Asparagus* from the Huétor-Tájar Population Variety. *J. Agric. Food Chem.* **2007**, 55 (24), 10028-10035.
79. Jiménez-Sánchez, C.; Pedregosa, F.; Borrás-Linares, I.; Lozano-Sánchez, J.; Segura-Carretero, A., Identification of Bioactive Compounds of *Asparagus officinalis* L.: Permutation Test Allows Differentiation among "Triguero" and Hybrid Green Varieties. *Molecules (Basel, Switzerland)*. **2021**, 26 (6), 1640.
80. Fuentes-Alventosa, J. M.; Jaramillo, S.; Rodríguez-Gutiérrez, G.; Cermeño, P.; Espejo, J. A.; Jiménez-Araujo, A.; Guillén-Bejarano, R.; Fernández-Bolaños, J.; Rodríguez-Arcos, R., Flavonoid Profile of Green *Asparagus* Genotypes. *J. Agric. Food Chem.* **2008**, 56 (16), 6977-6984.
81. Zhang, M.; Zhao, G.; Zhang, G.; Wei, X.; Shen, M.; Liu, L.; Ding, X.; Liu, Y., A targeted analysis of flavonoids in asparagus using the UPLC-MS technique. *Czech J. Food Sci.* **2020**, 38, 77-83.
82. gv, J.; Kandikattu, H. K.; K, K.; P, R.; Khanum, F., LC-ESI-MS/MS analysis of *Asparagus racemosus* Willd. roots and its protective effects against t-BHP induced oxidative stress in rats. *Ind Crops Prod.* **2015**, 78, 102-109.
83. Kamat, J. P.; Bloor, K. K.; Devasagayam, T. P. A.; Venkatachalam, S. R., Antioxidant properties of *Asparagus racemosus* against damage induced by γ -radiation in rat liver mitochondria. *J. Ethnopharmacol.* **2000**, 71 (3), 425-435.
84. Karuna, D. S.; Dey, P.; Das, S.; Kundu, A.; Bhakta, T., In vitro antioxidant activities of root extract of *Asparagus racemosus* Linn. *J. Tradit. Complement. Med.* **2018**, 8 (1), 60-65.
85. Lei, L.; Ou, L.; Yu, X., The antioxidant effect of *Asparagus cochinchinensis* (Lour.) Merr. shoot in d-galactose induced mice aging model and in vitro. *J Chin Med Assoc.* **2016**, 79 (4), 205-211.
86. Hamdi, A.; Jaramillo-Carmona, S.; Srairi Beji, R.; Tej, R.; Zaoui, S.; Rodríguez-Arcos, R.; Jiménez-Araujo, A.; Kasri, M.; Lachaal, M.; Karray Bouraoui, N.; Guillén-Bejarano, R., The phytochemical and bioactivity profiles of wild *Asparagus albus* L. plant. *Int. Food Res. J.* **2017**, 99, 720-729.
87. Olivier, M. T.; Muganza, F. M.; Shai, L. J.; Gololo, S. S.; Nemitavhanani, L. D., Phytochemical screening, antioxidant and antibacterial activities of ethanol extracts of *Asparagus suaveolens* aerial parts. *S. Afr. J. Bot.* **2017**, 108, 41-46.
88. Adouni, K.; Zouaoui, O.; Chahdoura, H.; Thouri, A.; Lamine, J. B.; Santos-Buelga, C.; González-Paramás, A. M.; Maggi, F.; Mosbah, H.; Haouas, Z. J. J. o. F. F., In vitro antioxidant activity, α -glucosidase inhibitory potential and in vivo protective effect of *Asparagus stipularis* Forssk aqueous extract against high-fructose diet-induced metabolic syndrome in rats. *J. Funct. Foods.* **2018**, 47, 521-530.
89. Nwafor, P. A.; Okwuasaba, F. K., Anti-nociceptive and anti-inflammatory effects of methanolic extract of *Asparagus pubescens* root in rodents. *J. Ethnopharmacol.* **2003**, 84 (2), 125-129.
90. Lee, H. A.; Koh, E. K.; Sung, J. E.; Kim, J. E.; Song, S. H.; Kim, D. S.; Son, H. J.; Lee, C. Y.; Lee, H. S.; Bae, C. J.; Hwang, D. Y., Ethyl acetate extract from *Asparagus cochinchinensis* exerts anti-inflammatory effects in LPS-stimulated RAW264.7 macrophage cells by regulating COX-2/iNOS, inflammatory cytokine expression, MAP kinase pathways, the cell cycle and antioxidant activity. *Mol. Med. Rep.* **2017**, 15 (4), 1613-1623.
91. Sung, J.-E.; Lee, H.-A.; Kim, J.-E.; Yun, W.-B.; An, B.-S.; Yang, S.-Y.; Kim, D.-S.; Lee, C.-Y.; Lee, H.-S.; Bae, C.-J.; Hwang, D.-Y., Saponin-enriched extract of *Asparagus cochinchinensis* alleviates airway inflammation and remodeling in ovalbumin-induced asthma model. *Int. J. Mol. Med.* **2017**, 40 (5), 1365-1376.
92. Hildah Mfengwana, P.-M.-A.; Sitheni Mashele, S. In *Medicinal Properties of Selected Asparagus Species: A Review*, 2019.
93. Onlom, C.; Khanthawong, S.; Waranuch, N.; Ingkaninan, K., In vitro anti-Malassezia activity and potential use in anti-dandruff formulation of *Asparagus racemosus*. *Int. J. Cosmet. Sci.* **2014**, 36 (1), 74-78.
94. Rosado-Álvarez, C.; Molinero-Ruiz, L.; Rodríguez-Arcos, R.; Basallote-Ureba, M. J.,

- Antifungal activity of asparagus extracts against phytopathogenic *Fusarium oxysporum*. *Sci. Hortic.* **2014**, *171*, 51-57.
95. Sairam, K.; Priyambada, S.; Aryya, N. C.; Goel, R. K., Gastroduodenal ulcer protective activity of *Asparagus racemosus*: an experimental, biochemical and histological study. *J. Ethnopharmacol.* **2003**, *86* (1), 1-10.
96. Farah, P., Antifungal Activity of Some Ethnomedicinally Important Tuberous Plants of Family Liliaceae. *Natl Acad Sci Lett.* **2020**, v. *43* (no. 1), pp. 93-97-2020 v.43 no.1.
97. Mandal, S. C.; Nandy, A.; Pal, M.; Saha, B. P., Evaluation of antibacterial activity of *Asparagus racemosus* Willd. root. *Phytother Res.* **2000**, *14* (2), 118-119.
98. Asnaashari, S.; Dastmalchi, S.; Javadzadeh, Y., Gastroprotective effects of herbal medicines (roots). *Int. J. Food Prop.* **2018**, *21* (1), 902-920.
99. Nwafor, P. A.; Okwuasaba, F. K.; Binda, L. G., Antidiarrhoeal and antiulcerogenic effects of methanolic extract of *Asparagus pubescens* root in rats. *J. Ethnopharmacol.* **2000**, *72* (3), 421-427.
100. Amina, M.; Al Musayeib, N. M.; Alarfaj, N. A.; El-Tohamy, M. F.; Al-Hamoud, G. A., Antibacterial and Immunomodulatory Potentials of Biosynthesized Ag, Au, Ag-Au Bimetallic Alloy Nanoparticles Using the *Asparagus racemosus* Root Extract. *Nanomaterials* **2020**, *10* (12), 2453.
101. Ajazuddin; Saraf, S., Applications of novel drug delivery system for herbal formulations. *Fitoterapia.* **2010**, *81* (7), 680-689.
102. Marslin, G.; Sheeba, C. J.; Franklin, G., Nanoparticles Alter Secondary Metabolism in Plants via ROS Burst. *Front. Plant Sci.* **2017**, *8*.
103. Vijay Kumar, P. P. N.; Kalyani, R. L.; Veerla, S. C.; Kollu, P.; Shameem, U.; Pammi, S. V. N., Biogenic synthesis of stable silver nanoparticles via *Asparagus racemosus* root extract and their antibacterial efficacy towards human and fish bacterial pathogens. *Mater. Res. Express.* **2019**, *6* (10), 104008.
104. Khanra, K.; Panja, S.; Choudhuri, I.; Anindita, C.; Bhattacharyya, N., Bactericidal and Cytotoxic Properties of Silver Nanoparticle Synthesized from Root Extract of *Asparagus racemosus*. *Nano Biomed Eng.* **2016**, *8*, 39-46.
105. Satyanarayana, B. M.; Reddy, N. V.; Kommula, S. k. R.; Rao, J. V., Biogenesis of silver nanoparticles using leaf extracts of *Asparagus racemosus* and *Sophora interrupta*: structure characterization, antibacterial and anticancer studies. *SN Appl. Sci.* **2020**, *2* (11), 1857.
106. Kote, J.; Kadam, A.; Patil, S.; Mane, R., Green functionalized silver nanoparticles With significantly enhanced Antimycobactericidal and Cytotoxicity Performances of *Asparagus racemosus* Linn. *Int. J. Eng. Sci. Technol.* **2016**, *3*, 12-26.
107. Mani, R. K.; Ahmed, S. S.; Babu, A.; Delna, G.; Jacob, L. P.; S, V.; Bharathi, B., Invitro evaluation of silver nanoparticle of *Asparagus racemosus* for it's anti diabetic activity. *UPI j. pharm. med. health sci.* **2021**, *4* (3), 01-05.
108. Varaprasad, T.; Govindh, B.; Rao, B. V., Green Synthesized Cobalt Nanoparticles using *Asparagus racemosus* root Extract & Evaluation of Antibacterial activity. *Int. J. Chemtech Res.* **2017**, *10*, 339-345.
109. Tripathi, D.; Modi, A.; Smita, S. S.; Narayan, G.; Pandey-Rai, S., Biomedical potential of green synthesized silver nanoparticles from root extract of *Asparagus officinalis*. *J. Plant Biochem. Biotechnol.* **2022**, *31* (1), 213-218.
110. Thakur, S.; Sharma, S.; Thakur, S.; Rai, R., Green Synthesis of Copper Nano-Particles Using *Asparagus adscendens* Roxb. Root and Leaf Extract and Their Antimicrobial Activities. *Int. j. curr. microbiol. appl. sci.* **2018**, *7*, 683-694.
111. Thakur, R.; Sharma, S., Study the antibacterial activity of copper nanoparticles synthesized using herbal plants leaf extracts. *Int. J. Biotech. Res (IJBTR).* **2014**, *4*, 21-34.
112. Pallela, P. N. V. K.; Ummey, S.; Ruddaraju, L. K.; Kollu, P.; Khan, S.; Pammi, S. V. N., Antibacterial activity assessment and characterization of green synthesized CuO nano rods using *Asparagus racemosus* roots extract. *SN Appl. Sci.* **2019**, *1* (5), 421.
113. Plangsombat, N.; Rungsardthong, K.; Kongkaneramt, L.; Waranuch, N.; Sarisuta, N., Anti-inflammatory activity of liposomes of *Asparagus racemosus* root extracts prepared by various methods. *Exp. Ther. Med.* **2016**, *12* (4), 2790-2796.
114. Adouni, K.; Júlio, A.; Santos-Buelga, C.; González-Paramás, A. M.; Filipe, P.; Rijo, P.; Costa Lima, S. A.; Reis, S.; Fernandes, Â.; Ferreira, I. C. F. R.; Fernández-Ruiz, V.; Morales, P.; Flamini, G.; Achour, L.; Fonte, P., Roots and rhizomes of wild *Asparagus*: Nutritional composition, bioactivity and nanoencapsulation of the most potent extract. *Food Biosci.* **2022**, *45*, 101334.