



Effects of Phytohormones on Reversing the Inhibitory Action of Aflatoxin B1 on the Growth of Maize Seeds (*Zea mays* L.)

Ganjendra Prasad¹, Vijendra Kumar Mishra^{2*} and Nitu Kumari¹

¹Plant Physiology and Mycotoxin Laboratory, Department of Botany, L. N. Mithila University, Kameshwarnagar, Darbhanga-846004, Bihar, India.

²Department of Biotechnology, L. N. Mithila University, Kameshwarnagar, Darbhanga-846004, Bihar, India.

Authors' contributions

This work was carried out in collaboration between all authors. Author GP designed the study, performed the statistical analysis, wrote the protocol, data interpretation and wrote the first draft of the manuscript. Author VKM managed the analyses of the study, edited and finalized the draft into manuscript format, literature searches and correspondence of the manuscript. Author NK managed the technical searches. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IJPSS/2018/40097

Editor(s):

(1) Francisco Cruz-Sosa, Department of Biotechnology, Metropolitan Autonomous University Iztapalapa Campus, Av. San Rafael Atlixco 186 México City, México.

Reviewers:

- (1) İrfan Terzi, University of Dumlupınar, Turkey.
(2) Akinyosoye, Solomon Tayo, Institute of Agricultural Research and Training, Obafemi Awolowo University, Nigeria.
(3) S. O. Olubodun, University of Benin, Nigeria.
(4) Mekbib Sissay Bekele, National University of Lesotho, Lesotho.

Complete Peer review History: <http://www.sciencedomain.org/review-history/24475>

Short Communication

Received 14th February 2018
Accepted 30th April 2018
Published 5th May 2018

ABSTRACT

The present study is aimed to investigate the effect of phytohormones namely gibberellic acid (GA₃) and kinetin on aflatoxins B1 treated maize seeds var madhuri-01. Maize seeds were treated with aflatoxins B1 separately and in combination of phytohormones at a concentration of 2.0 ppm each. Reversal of inhibitory effect of aflatoxin B1 on the seed germination, root, shoot length, chlorophyll a, b and carotenoid of maize seed in presence of phytohormone was observed. The inhibitory effect of aflatoxins B1, on seed germination, root length, shoot length, chlorophyll a, chlorophyll b and

*Corresponding author: E-mail: vijendrkmishra@gmail.com, drvijendrakumarmishra@gmail.com;

carotenoid content were 78.57%, 65.03%, 63%, 74%, 72% and 85%, respectively. However, when the maize seeds were treated in combination of aflatoxins B1 with kinetin and aflatoxin B1 with GA₃, reversal of the inhibitory effect of aflatoxin B1 was observed to great extent. Taken together, the reversal of phytoinhibitory effect of aflatoxin B1 by GA₃ and kinetin could be of agricultural significance.

Keywords: Chlorophyll; carotenoids; aflatoxin-B₁; phytohormones and *Zea mays* L.

1. INTRODUCTION

Maize is one of the largest and most important cereal crops produced in the world. In 2012, the total production of maize was 872 million metric tons. Maize (also known as corn) is a domesticated cereal grain that has been grown as food and animal feed for tens of thousands of years. Maize (*Zea mays* L.) is an important crop for human consumption, particularly in developing countries, where this cereal can represent up to 65% of the total calories and 53% of the protein intake [1]. Maize is an important cereal crop and consumed by almost entire population in India in various ways. It is cultivated in all the three seasons' viz. Kharif (Monsoon), winter and summer in India.

Inhibitory effect of aflatoxin B1 on various cereal crops has been well documented [2,3,4,5]. The inhibitory effect of mycotoxins results in severe losses of maize productivity. The previous studies showed that the role of phytohormones like GA₃ and kinetin as anti-toxicants as well as the phytostimulatory agents. Adverse effect of aflatoxins B1 on physiological process of *Zea mays*, *Vigna radiate*, such as reduction in seed germination, seedling growth has been well documented [6,7]. Aflatoxins B1 is known to interfere with the synthesis of GA₃ which stimulates the activities of lipase and α-amylase enzyme, primarily required during seed germination and seedling growth [8]. Literature survey shows that GA₃ plays a key role in seed dormancy breaking, seedling growth and induce seed germination in soyabean seeds [9]. Brown et. al. (1986) [10] proposed that GA₃ acted the most active group of hormones in the breaking of dormancy in *Protea Compacta R-BR*, cytokinin (kinetin) also act activity in stimulating of radicles growth in green gram [2,3]. Since only the few reports are available about the impact of phytohormones on mycotoxins treated seeds, the present work was designed to find out if the additional dose of GA₃, and kinetin can reverse the inhibitory effect of aflatoxins -B₁ on germination and growth of maize seedlings.

2. MATERIALS AND METHODS

Seeds of maize var. Madhuri-01 were obtained from Dayal Traders Manures and seed storage house, Kadirabad, Darbhanga, Bihar, India. aflatoxins-B₁ and phytohormones such as GA₃ and kinetin were obtained from the local Scientific Stores, Darbhanga of sigma, USA. The concentration of aflatoxins-B1, GA₃ and kinetin used was 2.0 ppm only. Since the effects of 5 different concentrations of aflatoxins B1 (0.1, 0.25, 0.50, 1.0 and 2.0 ppm) on seed germination, seedling growth, chlorophylls and carotenoids synthesis of maize seeds were studied during Ph.D. work [11]. The maximum inhibition in above parameters were recorded at 2.0 ppm of aflatoxins B1 and hence, already established concentration of aflatoxin B1 (2.0 ppm) was used in the present study [11]. Stock solution was initially prepared in 1.0 mL ethanol from which the dilution (2.0 gm⁻³ change to mL) was made in sterile double distilled water solutions of aflatoxins B1 toxin and phytohormones such as GA₃ and kinetin were mixed separately and in combination also, at a ratio of 1:1 (v/v), at 2.0 ppm to investigate their combined effects. Maize seeds were soaked in double distilled water for 1 h and subsequently in different combinations of these toxins and phytohormones for 20 h. One hundred healthy maize seeds were selected for each treatment in triplicate set. The soaked seeds were subsequently placed on moist blotting paper and incubated in seed germinator at 18±2 °C. Seed germination index (GI) was calculated after 5-days of incubation according to the formula as given below.

$$GI = \frac{\text{No. of germinated seeds}}{\text{No. of seed observed}} \times 100$$

Radical and plumule length was measured in centimeter after 7-days of growth. Chlorophyll-a, b and carotenoid contents were estimated according to the method described by Arnon (1949) [12] and Davis (1976) [13], respectively. The data were analyzed statistically that is "t-

test” for seed germination and F-test for seedling growth as well as chlorophyll contents. Statistical calculations were carried out using ANOVA test [14].

3. RESULTS AND DISCUSSION

The effect of GA₃ and kinetin on aflatoxins B1 treated maize seed grown seedling germination, root and shoot length is represented in Table 1 and Table 2. The maximum inhibition in seed germination was 78.57% in presence of aflatoxins B1 only. However, minimum inhibition was observed with kinetin (19.38%) and GA₃ (10.20% in combination with aflatoxins B1. It was observed that aflatoxins B1 drastically inhibited seed germination, root and shoot length when applied separately. On the other hand, these parameters increased when treated in combination with GA₃ and kinetin phytohormone in presence of aflatoxins B1. Students t-test Para analysis showed significant effect (P<0.01) on seed germination due to aflatoxins B1 and with phytohormones treatments. The lowest inhibition recorded in root length and shoot length was 8.79% and 19.00% in GA₃ treated seeds, respectively. However, due to kinetin treatment the lowest inhibition in root and shoot length was 11.55% and 21.00%, respectively. The highest inhibition in root and shoot length was 65.03% and 63% respectively, in presence of aflatoxins B1 only. However, the treatment of aflatoxins-B1 with kinetin exhibited little bit greater inhibition as compared to the aflatoxins-B1 with GA₃ treatments. The longest root length was observed in GA₃ (12.11 cm) followed by kinetin (11.76 cm) treated maize seeds. However, the shortest root length was observed in aflatoxins B1 treated seeds followed by B1 with kinetin (8.92 cm) and aflatoxin B1 with GA₃ (8.65 cm) seeds of maize. The longest shoot length was

observed in GA₃ (8.89 cm) followed by kinetin (8.76 cm) treated maize seeds. However, the shortest shoot length was observed in aflatoxins B1 treated B1 with Kinetin (4.83 cm) and B1with GA₃ (4.94 cm) seeds of maize. It was observed that GA₃ and kinetin reverses the toxic and inhibitory effect of aflatoxins-B1 as shown in Table 2. The effect of aflatoxins B1 and phytohormones on the chlorophyll a, b and carotenoid is represented in Table 3. It was observed that aflatoxins B1 drastically inhibited chlorophyll a, b and carotenoid. On the other hand these parameters reversed when treated in combination with GA₃ and kinetin phytohormone in presence of aflatoxins B1, separately. The lowest inhibition was recorded in Chl-a, Chl-b, Total Chl (a+b) and carotenoids were 16.00%, 10.00%, 11.00% and 28.00% due to GA₃ treatment. However, due to kinetin treatment the inhibition in Chl-a, Chl-b, total Chl (a+b) and carotenoids was 18.00%, 20.00%, 19.00% and 35.00% in maize seeds. Aflatoxin B1 with kinetin showed more inhibition in comparison to the toxin with GA₃. The drastic reduction as well as enhancement of total chlorophyll and carotenoids content in aflatoxins-B1 treated seed was noticed due to stimulatory effects of GA₃ and kinetin. GA₃ also reverses the inhibition caused by the maximum concentration of aflatoxins-B1 in Cotton seeds. Our results are like the other reports [8]. Singh et. al. (1991) [15] also recorded the similar inhibition reversed effect of GA₃ caused due to aflatoxins B1 in the seeds of chick pea and pigeon pea [16]. There are only few reports are available on the stimulating effect of GA₃ on the chlorophyll synthesis [17]. Above result reveals that GA₃ and kinetin play crucial role in reversal of toxic effect of aflatoxins B1 treated seeds by breaking dormancy of seeds, which warrants further investigations.

Table 1. Impact of GA₃ and kinetin on aflatoxin B1 treated maize seeds #

Observations	Control	GA ₃	Kinetin	aflatoxin B ₁	aflatoxin B ₁ + GA ₃	Aflatoxin B ₁ + Kinetin
Seed germination (Index X ± S.E).	98 ± 0.47	100 ± 1	100 ± 1	21 ± 0.57	88 ± 0.47	79 ± 0.74
% inhibition	-	-	-	78.57%	10.20%	19.38%
“t” Difference with Control	-	-2.00	-2.00	13.50	22.27	27.02
% Inhibition	-	-2.00	-2.00	78.57	10.20	19.38

p (0.01) (t – test), Significant at all the treatments and combinations of toxins and phytohormones

Table 2. Effect on root and shoot length of phytohormone GA₃ and Kinetin on aflatoxin B₁ treated maize seeds

Observations	Root length (cm)			Shoot length (cm)		
	X ± S.E	Difference with control	% inhibition	X ± S.E.	Difference with control	% Inhibition
Control	9.78 ± 0.12	–	–	6.15 ± 0.24	–	–
GA ₃	12.11 ± 1.00	2.33	– 23.82	8.89 ± 0.08	– 2.74	– 44.55
Kinetin	11.76 ± 0.15	0.35	– 20.24	8.76 ± 0.09	– 2.61	– 42.00
Aflatoxin-B ₁	3.42 ± 0.09	8.69	65.03	2.25 ± 0.08	3.90	63.00
Aflatoxin-B ₁ + GA ₃	8.92 ± 0.07	3.19	8.79	4.94 ± 0.12	1.21	19.00
Aflatoxin-B ₁ + Kinetin	8.65 ± 0.04	3.46	11.55	4.83 ± 0.12	1.32	21.00
LSDO ₁	–	0.33	3.80	–	0.42	14.62
LSDO ₅	–	0.23	2.71	–	0.29	10.41

* Non significant at 1 and 5% level

** Non Significant at only 5% level, all other values are significant at both 1 and 5% level

Table 3. Effects of GA₃ and kinetin on aflatoxin B₁ treated chlophylls carotenoid contents of Maize seeds #

Observations (Concen.)	%Chl-a X ± S.E.	Difference with control	% inhibition	%chl – b X ± S.E.	Difference with control	% inhibition	Total Chl (a+b)	% inhibition	% Carotenoids	Difference with control	% inhibition
Control	0.880 ± 0.073	–	–	0.328 ± 0.159	–	–	1.216 ± 0.014	–	0.014 ± 0.023	–	–
GA ₃	0.979 ± 0.039	–0.099	– 11.25	1.159 ± 0.014	–1.262	–61.58	1.571 ± 0.029	–24.00	0.027 ± 0.023	–0.013	–92.00
Kinitin	0.926 ± 0.025	0.046	–4.60	1.488 ± 0.019	–1.160	–51.21	1.422 ± 0.029	16.00	0.026 ± 0.014	–0.012	–85.00
aflatoxins-B ₁	0.226 ± 0.023	0.654	74.00	0.274 ± 0.023	0.054	72.00	0.316 ± 0.014	74.00	0.002 ± 0.014	0.012	85.00
aflatoxins-B ₁ + GA ₃	0.737 ± 0.039	0.143	16.00	0.293 ± 0.053	0.035	10.00	1.081 ± 0.014	11.00	0.010 ± 0.023	0.004	28.00
aflatoxins-B ₁ + Kinitin	0.715 ± 0.039	0.165	18.00	0.261 ± 0.014	0.067	20.00	0.980 ± 0.014	19.00	0.009 ± 0.014	0.005	35.00

#% = mg/100mg

Significant at 1 & 5% level

4. CONCLUSION

Application of phytohormones like GA₃ and Kinetin could significantly reverse the inhibitory effect of aflatoxin B1 of maize seedlings. That could be of agricultural importance and warrants further investigations.

ACKNOWLEDGEMENTS

The authors are thankful to the Prof. and Head, University Department of Botany, L.N.M.U, Darbhanga for making use of laboratory facilities and thanks to the CSIR-I for financial assistance.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Manal MAR. Improvement the production of maize (*Zea Mays L.*) crop by using particle bombardment. international conference on biological, civil and environmental engineering (BCEE-2015) Feb. 3-4, Bali (Indonesia); 2015.
2. Cavusoglu AM, Sulusoglu. The effects of exogenous gibberellin on seed germination of the fruit species. DERLEME. 2015;8(1): 06-09.
3. Sakdeo BM. Physiological effect of seed treatments with kinetin on seedling growth under laboratory and field conditions in Green gram. Int. J. Appl. Res. 2016;2(7): 384-387.
4. Dilip WS, Singh D, Moharana D, Rout S, Patra SS. Effect of gibberellic acid (GA) different concentrations at different time intervals on seed germination and seedling growth of Rangpur lime. J. Agroeco. Nat. Resource Management. 2017;4(2): 157-165.
5. Reddy KRN, Raghavender CR, Reddy BN, Salleh B. Biological control of *Aspergillus flavus* growth and subsequent aflatoxin B1 production in sorghum grains. African J. of Biotech. 2010;9:4247-4250.
6. Bilgrami KS, Sinha KK, Masood A, Rahman MF. The effect of T-2 toxin on seed germination and seedling growth of mung (*Vigna radiata L.*) Nat. Acad. Sci. Litt. 1991;14:363-365.
7. Sinha KK, Punam Kumari. Some physiological abnormalities induced by aflatoxin B₁ in mung seeds (*Vigna radiata* variety pusa Baishakhi). Mycopathologia, 1990;1(10):77-79.
8. Chatterjee D. Inhibitory effects of aflatoxin B1 on amylase of maize seeds. Lters in Applied Microbiology. 1988;7(1): 9-11.
9. Tiwari RP, Bhalla TC, Saini SS, et al. Mechanism of action of aflatoxin B₁ J Biosci. 1986;10(1):145-155.
10. Brown NAC, Van Staden J, Jacobs G. Germination of achenes of *Leucospermum cordifolium*. Acta Hort. 1986;185:53-59.
11. Prasad G. Effects of aflatoxin B1 and citrin on physiological and biochemical process of maize seeds. Ph.D. thesis T.M.B.U. Bhagalpur; 1992.
12. Aronon DI. Copper enzymes in isolated chloroplast polyphenol oxidase in beta vulgaris – Plant Physio. 1949;24:1-15.
13. Davis BH. Carotenoids in chemistry and biochemistry of plant pigments, (Eds. T.W. Goodwin), Academic press, London. 1976;2:38-165.
14. Dospekhov BA. Field experimentation, statistical procedures mir, Moscow; 1984.
15. Singh PL, Ahmad SK, Bhagat S. In Mycotoxin incidence and human health (eds. K.S. Bilgrami and K.K. Sinha) Allied Press, Bhagalpur. 1991;83-87.
16. Kabar K. Comparison of kinetin and gibberellic acid effects on seed germination under saline conditions. Phytion (Austria). 1990;30(2):291-298.
17. Singh MK, Sinha KK. Levels of aflatoxin B1 production in seeds of some selected varieties of paddy and their relation with total starch, amylose and amylopectin contents. International Journal of Life Science Biotechnology and Pharma Research. 2013; 2(2):221-226.

© 2018 Prasad et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<http://www.sciencedomain.org/review-history/24475>