# academicJournals

Vol. 11(22), pp. 945-954, 14 June, 2017 DOI: 10.5897/AJMR2016.8340 Article Number: 93FED9664703 ISSN 1996-0808 Copyright © 2017 Author(s) retain the copyright of this article http://www.academicjournals.org/AJMR

**African Journal of Microbiology Research** 

Full Length Research Paper

# Biochemical and molecular identification of newly isolated pigmented bacterium and improved production of biosurfactant

Helvia W. Casullo Araújo<sup>1</sup>, Rosileide F. S. Andrade<sup>2,5</sup>, Dayana Montero-Rodríguez<sup>3,5</sup>, Vanessa Pimentel Santos<sup>4,5</sup>, Patrícia C. V. Souza Maia<sup>4,5</sup>, Carlos F. B. Costa Filho<sup>4,5</sup>, Carlos A. Alves da Silva<sup>5</sup> and Galba Maria Campos-Takaki<sup>5\*</sup>

<sup>1</sup>Department of Chemistry, State University of Paraíba, 58429-500, Campina Grande, Paraíba, Brazil. <sup>2</sup>National Post-Doctorate Program-CAPES, Catholic University of Pernambuco, 50050-900 Recife-PE, Brazil. <sup>3</sup>Center for Biological Sciences, Federal University of Pernambuco, 50670-901 Recife, Pernambuco, Brazil. <sup>4</sup>Master in Development of Environmental Processes, Catholic University of Pernambuco, 50.050-900 Recife, Pernambuco, Brazil.

<sup>5</sup>Nucleus of Research in Environmental Sciences and Biotechnology, Catholic University of Pernambuco, 50050-590, Recife, Pernambuco, Brazil.

Received 18 October 2016, Accepted 17 November, 2016

A novel bacterium pigmented isolate from Caatinga soil was characterized by biochemical and molecular assays, as well as, by rep-polymerase chain reaction (PCR) and 16S rDNA sequencing, and was identified as *Serratia marcescens* based on 99% of similarity. The identity of the sequences were compared by pairs of critical species of *S. marcescens* found in National Center for Biotechnology Information (NCBI) with a 96% homology of the isolated species to species database. The wild strain was able to produce biosurfactant (BS) using cassava (*Manihot esculenta* Crantz) wastewater, with addition of lactose, and corn oil, according to full factorial design 2<sup>3</sup>, at 28°C, and static condition. The net liquid metabolic reduced the surface tension of the water from 70 to 30.60 mN/m during the stationary phase (assay eight constituted by 6.0% cassava wastewater, 1.0% lactose and 7.5% corn oil), and produced emulsifier agent (4.012 UEA) at the same condition. This study identified the pigmented bacterium as new strain of *S. marcescens*, and showed it has potential to promote both emulsions formation and surface tension reduction in cassava wastewater (6.0%), lactose (1.0%) and corn oil (7.5%) proved as an alternative economical medium for commercial biosurfactant processes.

Key words: Serratia marcescens, tensio-active, bioemulsifier, cassava wastewater.

# INTRODUCTION

The semi-arid region of Northeast of Brazil comprises

approximately 969.589.4 of km<sup>2</sup> (Sousa et al., 2015). The

soil composition particularly those under native vegetation for semi-arid is latosols and luvisols under native vegetation for representative sequences from Pernambuco (Sá et al., 2015). This region has a dry tropical deciduous vegetation, constituted by small trees, bushes and grasses, xerophiles, where deciduous plants are the mainly vegetation. It is also characterized by high level of insolation, high temperature, scanty hydric resources and scarce rains, which often cause long periods of drought. In addition, all these factors are usually unfavorable for microbial growth in the soil; however, bacteria constitute the principal group of decomposers responsible for carbon recycling (Solanki and Arora, 2015).

The surfactants in its current use are chemically derived from petroleum; however, interest in microbial surfactants has been applied steadily as surface-active biomolecules that are produced by a variety of microorganisms. The use of low cost substrate can allow a reduction of more than 50% of the final costs of biosurfactant production (Brumano et al., 2016). In this study, cassava (Manihot esculenta Crantz) wastewater effluent was used as substrate. The tubercle has a great economic importance to several cities of Northeastern of Brazil, where its consumption (in terms of carbohydrate content) exceeds those of other crops. However, cassava contain a large amount of cyanogenic glucosides, which can be hydrolyzed to hydrocyanic acid (HCN) by the endogenous enzymes, when the plant tissue is damaged during harvesting, processing or other mechanical processes during the preparations. In this case, the effluent from cassava industrial processing is responsible for the largest environmental impact (Olaifa et al., 2015).

The aim of this study was to perform the conventional and molecular identification of the pigmented bacterium isolated from semi-arid soil, as well as to evaluate the potential of the new microorganism to biosurfactant production using cassava wastewater as economic substrate.

# MATERIALS AND METHODS

#### Microorganism

The new pigmented bacterium was isolated from Caatinga soil cultured with banana plants, and was deposited in the culture collection of the Catholic University of Pernambuco – UCP. After identification, the strain was registered in World Federation for Culture Collection - WFCC. The strain was maintained on nutrient agar at 5°C.

#### Substrate

The cassava wastewater was used as media for the production of the biosurfactant, and was obtained from industry food at municipal district of Carnaíba – Pernambuco, and the chemical composition as described by Ponte (1992).

#### Biochemical and molecular strain identification

The morphological and biochemical characteristic of the isolate bacterium was used for identification tests according to Bergey's Manual of Systematic Bacteriology (Gorlach-Lira and Coutinho, 2007) and the Manual of Methods for General Bacteriology (Holt et al., 1994).

identification, For the primers FD1 (5'AGAGTTTGATCCTGGCTCAG3') and RD1 (5'AAGGAGGTGATCCAGCC3') were used to amplify the 16S rDNA fragments. The PCR reaction mixture contained 50 ng genomic DNA (Sambrook and Russell, 2001). Sequences were compared with the ones in the databanks of the National Center Biotechnology Information, NCBI for site (http://www.ncbi.nlm.nih.gov/blast), using the program BlastX (Altschul et al., 1997). The sequences were aligned with the software ClustalW (http://www.ebi.ac.uk) (Thompson et al., 1984). Evolutionary trees for the data set were inferred by the method of Saitou and Nei (1987), and using the neighbor-joining program MEGA (Kumer et al., 1993). The stability of relationships was assessed by performing bootstrap analyses of the neighbor-joining data based on 1.000 resembling.

### Medium and culture conditions

The pre-inoculums was prepared by transferring the new strain S. *marcescens* grown in 100 mL of Luria Bertani (yeast extract 5 g, glucose 5 g, NaCl 5 g per liter), incubated at 28°C, and shacked by orbital shaker at 150 rpm during 17 h. Then, Erlenmeyer's flasks of 500 mL capacity containing 200 mL of the media according to the factorial design (Table 3) were inoculated with 10% of  $10^{6}$  UFC/mL according to the factorial design (Table 3). The flasks were incubated at 28°C in static condition, during the 72 h. The liquid metabolic free of cells was obtained using filtration through a Millipore membrane with a pore size of 0.22 µm, and were determined pH, surface tension (ST) (mN/m) and emulsification activity (UAE) were determined.

#### Emulsifier activity test

The metabolic liquid free of cells was added to 2.0 mL of sodium acetate buffer 0.1 M (pH 3), with 1.0 mL of n-hexadecane, the mixture was vortexed for 2 min. After 10 min of rest, the formed emulsions formed were removed with the aid of the Pasteur pipette and transferred and read in spectrophotometer at the wavelength, 540 nm (Cirigliano and Carman, 1984).

\*Corresponding author. E-mail: galba\_takaki@yahoo.com.br. Tel: +51-81-21194044. Fax: +55-81-21194043.

Author(s) agree that this article remains permanently open access under the terms of the <u>Creative Commons Attribution</u> <u>License 4.0 International License</u>

**Table 1.** Level and factors applied to factorial design compound  $2^3$ .

Levels	-1	0	+1
Lactose (% w/v)	0.20	0.60	1.00
Cassava wastewater (% v/v)	1.00	3.50	6.00
Oil Corn (% v/v)	5.00	6.25	7.50

 Table 2. Characteristics of the tests and biochemical morphophysiologic new pigmented bacterium was isolated from Caatinga soil.

Tests	Bacterium	Tests	Bacterium	Tests	Bacterium
Gram	-	DNase	+	Lactose	-
Catalase	+	Adonitol	-	Maltose	+
Urease	-	Arabinose	-	Mannitol	+
Lysine	+	Dextrose	+	Rafinose	-
Motility	+	Galactose	+	Sorbitol	+
Indole	-	Inositol	+	Color	Pink/Red
Gas H <sub>2</sub> S	+	Sucrose	+	Glycose	+

Negative: -; Positive: +

#### Surface tension (ST) measurement

The cell free supernatant was determined in a tensiometer model Sigma 70 (KSV Instruments LTD - Finland), using the Du Nouy ring method. The reported values are the means of 3 measurements of surface tension (ST) from the media were used as control (Kuyukina et al., 2001).

#### Experimental factorial design

Production of biosurfactant was performed using a full  $2^3$  factorial design, augmented with a 4 central points, used to evaluate the impact of 3 independent factors related to lactose, cassava wastewater and corn oil, on the response variable surface tension (ST) and emulsifier activity (Table 1).

Analysis of data was carried out using the analysis of variance (ANOVA). The results were analyzed then examined to determine the main effects of all factors. The ANOVA was carried out to determine which factors are statistically significant. The data analyses and graphs, and all calculations were performed using the Statistical 7.0 software package, and significance of the results was tested at the ( $p \le 0.05$ ).

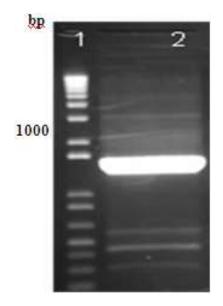
# **RESULTS AND DISCUSSION**

The Gram negative bacterium isolated from Caatinga banana tree soil showed red colonies on LB medium at 28°C during 24 h. The Rugai-Lysine test was employed (Pessoa and Silva, 1972) and catalase test showed positive reaction. The motility of gas production, indole, urease and sucrose showed negative reactions. The results identified the strain as *Enterobacteriacea* family, genus *Serratia* (Table 2).

The optimum growth conditions of *Serratia* were observed in pH 9 at temperatures of 20 to 30°C (Khanafari et al., 2006). The results obtained agree with those reported by Dundar et al. (2009). This method is widely used in bacterial systems. Apart from the morphology, staining can reveal certain properties of the bacteria which are essential for classifying it. Under this method, bacteria are classified into two groups: Gram positive and negative, the isolate examined was a Gram negative bacillus, which is alcohol acid resistant.

The sample also showed red-pink color when grown in Luria Bertani media at 28°C for 24 h. The Rugai-Lysine test was employed (Pessoa and Silva, 1972) and showed a positive reaction for lysine, motility and the production of gas, and a negative reaction for indole, urease and sucrose. Therefore, with this method, the examined strains can be identified as a genus *Serratia* sp. of the *Enterobacteriacea* species. The catalase test was positive for the sample, so the results of the morphological and biochemical analysis of the isolate can identify it as an *Enterobacteriacea* species of the genus *Serratia marcescens*, according to Holt et al. (1994).

The bacteria of the species *S. marcescens* may be identified in the laboratory through Gram-staining, although sometimes they can be observed without the use of laboratory equipment since at room temperature



**Figure 1.** Electrophoresis of PCR amplified 16S rDNA gene sequence of the DNA by the strain *S. marcescens*: lane 1 DNA model; lane 2 DNA UCP 1549.

they display pink to red pigmentation. *S. marcescens* displays an intense red pigmentation, prodigiosin by age of the colonies (Rastegari and Karbalaei-Heidari, 2016). To confirm the morphological and biochemical tests, molecular tests were also used. The DNA isolate was subjected to PCR reactions with primers specific for 16S rRNA region, which resulted in the amplification of a fragment of approximately 1200 pb (Figure 1). Genes of 16S rDNA have long been the standard for systematic molecular studies, as well as for the rapidly expanding fields of molecular diagnostics and microbial ecology.

These genes are universal multiple copies of 16S rDNA genes of the genome; thus making it abundant, easily detectable. Primers were designed to amplify fragments specific to the analysis of sequence. Gaps in 16S rRNA as a target for detection and identification are related to the fact, and they are often insufficient to discriminate the sequence information within the target 16S rRNA in order to distinguish among closely related species and strains (Ma et al., 2016).

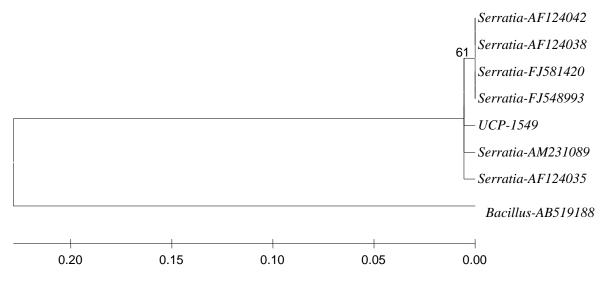
The isolated DNA was subjected to PCR reactions with specific primers for 16S rRNA, which resulted in the amplification of a fragment of approximately 1200 bp (Figure 1). The 16S rDNA have been used as standard for molecular systematic studies as well as the rapid expansion of the areas of molecular diagnostics and microbial ecology (Vierheilig et al., 2015).

The complete nucleotide sequence of isolate by 16S

rDNA gene, was compared by blast software specifically designed by the NCBI GenBank (BLAST/bl2seq) (GenBank). The identity of the sequences were compared by pairs of critical species *S. marcescens* found in NCBI/National Center for Biotechnology Information, a 96% homology of the isolated species to species database of NCBI was observed. The construction of the phylogenetic tree (Figure 2), in conjunction with the BLAST search, was gaining immense importance among researchers. This method relates the strain in question with those exhibiting a sequence homology of DNA and helps to overcome the possibility of errors by performing statistical analysis, such as testing the boot (Weisburg et al., 1991).

Two bodies can only be compared if they show similarity at least 80% in the primary sequences of their DNA. Indeed, hybrids showing 90-95% similarity in their sequences of nucleotide bases of DNA hybridization, showed 50% DNA (Weisburg et al., 1991). Most studies used this method, organisms that have a high homology (above 50%) have similar phenotypic characteristics. The correlation between phenotypic and genotypic properties led to the proposal that organisms with prokaryotic DNA homology values of less than 70% are considered as belonging to the same genomic species. The analysis of the 16S rDNA sequences confirmed the strain catalogued as UCP 1549 was closely related to S. marcescens based on 99% of similarity in their 16S rDNA sequences. The results indicate that amplified 16S rDNA analysis is a sample and reliable tool for the identification of bacterial species according to the literature (Zhu et al., 2007; Tariq and Prabakaran, 2011).

Cassava wastewater was a nutrient source for biosurfactant production by S. marcescens and the use of agroindustrial substrate manipueira could reduce the economic process of production. On the other hand, cassava processing is generally considered to contribute significantly to environmental pollution and aesthetic nuisance. In this study, the identified bacterium, S. marcescens demonstrated ability for a very fast growth and to convert cassava wastewater, lactose and corn oil are important nutrients to biosurfactant production detected by surface tension, emulsifier activity using a factorial design, after 72 h of growth, in the static condition. The best results observed were a significant surface reduction tension of water (70 mN/m to 30.60 mN/m) in condition 8 of the factorial design, with greater concentration of cassava wastewater (6.0%), lower level of lactose (1%), and higher level of corn oil (7.5%). At the same time, higher emulsifier activity in the assay 4 (cassava wastewater 6.0%, lower level of lactose 1%, and higher level of corn oil 5%) was observed (Table 3). According to the Pareto diagram (Figure 3A), the independent corn oil and lactose and interaction



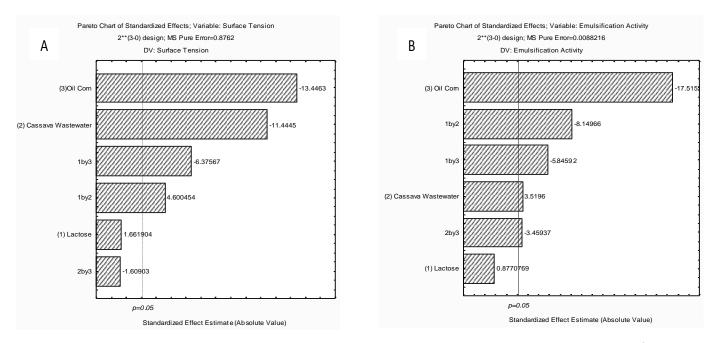
**Figure 2.** Phylogenetic dendrogram based on a comparison of 16S rDNA sequences for strain UCP 1549 among related bacteria. The tree was produced by using the maximum-likelihood algorithm and was calculated by using 1000 bases of the 16S.

	Parameters		Variables responses			
Assay	Lactose (%)	Cassava wastewater (%)	Oil Corn (%)	Surface tension(mN/m)	Emulsification activity (mN/m)	
1	0.2	1.0	5.0	47.60	3.102	
2	1.0	1.0	5.0	48.25	3.657	
3	0.2	6.0	5.0	36.42	3.674	
4	1.0	6.0	5.0	46.41	4.012	
5	0.2	1.0	7.5	42.36	2.124	
6	1.0	1.0	7.5	37.82	2.768	
7	0.2	6.0	7.5	32.30	3.102	
8	1.0	6.0	7.5	30.60	1.798	
9	0.6	3.5	6.25	33.21	3.476	
10	0.6	3.5	6.25	35.30	3.501	
11	0.6	3.5	6.25	34.61	3.600	
12	0.6	3.5	6.25	33.68	3.680	

**Table 3.** Biosurfactant production by *S. marcescens*: surface tension (ST) and emulsification activity using factorial design 2<sup>3</sup>, during 72 h, at 28°C as variables response.

lactose/corn oil favored positively the reduction of surface tension; however, the interaction lactose/cassava even considered significant at a confidence level within 5 % (p <0.05) and did not contribute to the reduction but was not statistically representative. The emulsification activity according to the Pareto diagram (Figure 3B) showed that cassava had no effect on emulsifying activity and corn oil as well as the interactions between lactose/cassava, lactose/corn oil, which was statistically more significant influence the emulsification activity. However, the emulsification activity was very promising for all conditions of the factorial design, with the highest emulsification activity in test 4 (4.012 UAE), and lowest in the assay 8 (UAE 1.798) (Table 2).

A study was carried out using *Serratia* sp with a reduction of the surface tension to 27.8 mN/m, after 72 h of cultivation (Montero-Rodríguez et al., 2015). Similar results with *S. marcescens* was observed by Ferraz et al. (2002) which demonstrated the ability for reduction of the surface tension to 29.75 mN/m by, but using orbital



**Figure 3.** Pareto chart of standardized effects of the net metabolic liquid from *S. marcescens* after 72 h of cultivation with 2<sup>3</sup> full factorial design. The point estimates the statistically significance (at p=0.05). (A) Surface tension; (B) Emulsification activity.

shaker. The obtained statistical model explained 85% of data variation and provide accreditations established of the experimental model. The Pareto chart (Figure 3A) reinforces the prediction model established by ANOVA. The statistical model of accuracy of surface tension as a function of the parameters was obtained by the following equation:

TS = 49.84 + 22.36 [Lactose] - 1.36 [Cassava] - 0.43 [Corn oil] + 1.53[lactose] [cassava] - 4.21[corn oil] [lactose] - 0.17 [cassava] [corn oil].

Araújo et al. (2009) described that the previous results of biosurfactant production by *S. marcescens* using as substrate lactose, corn oil and corn step liquor in substitution of cassava waste water and the reduction of the surface tension was 33.35 mN/m. Experimental factorial design conducted with *Serratia* sp. showed a reduction of the surface tension of the medium from 67.8 to 34.4 mN/m after 96 h of cultivation (Cunha et al., 2004).

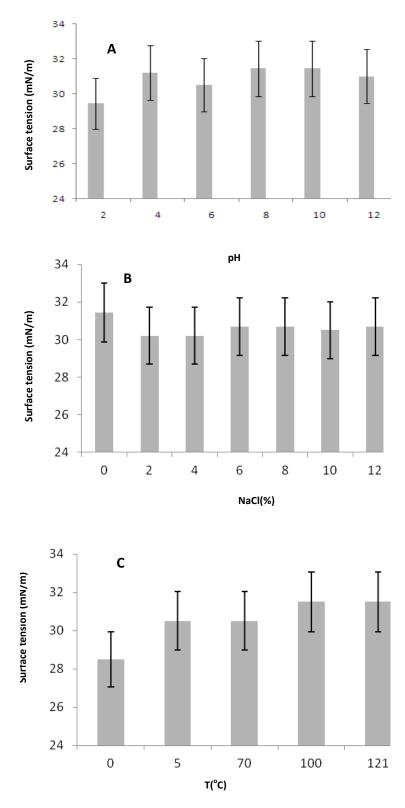
The stability of the biosurfactant was evaluated by the pH, the saline concentration and the temperature. Several studies described the of biosurfactants production optimization by culture media of low cost associated with industrial residue as insoluble substrates (Silva et al., 2010; Joshi et al., 2013; Ebadipour et al., 2016).

The biosurfactant produced by *S. marcescens* in the best condition of factorial design (assay 8) evaluated by surface tension of cell-free metabolic net showed stability under all different pH, NaCl concentrations and temperature.

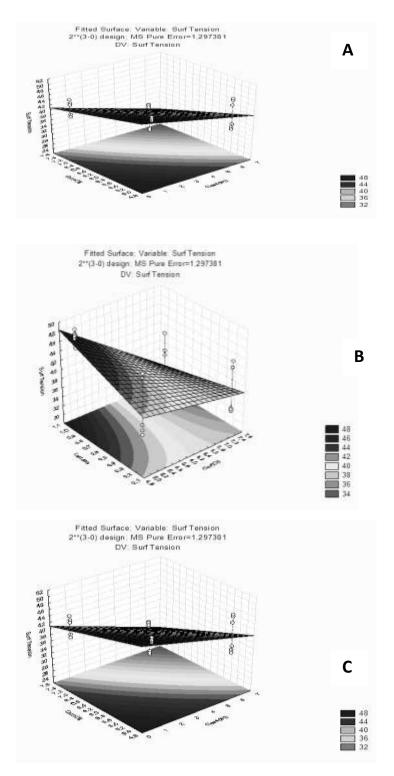
The stability of the produced biosurfactant produced was influenced in reducing surface tension by acid pH, salt concentration up to 6%, and the temperature 100°C (Figures 4A, B and C), respectively. However, there was no significant difference in the variation of surface tension, indicating its potential application in aquatic environments, because it is independent of the used concentration of NaCl and the biosurfactant remained effective in reducing the surface tension (Figure 4B). These results of the stability of the biosurfactant to 10% NaCl concentration, considering that conventional surfactants are inactivated by NaCl concentrations of 2 and 3%, as described by Rufino et al. (2007). The response surface graphs lactose/corn oil (Figure 5B), cassava/lactose (Figure 5A), have significant interactions, that is, cannot be explained by variations alone, because their curves were not parallel. Figure 5C showed the curves of levels of character parallel, and surface response indicated considerable interaction between the used media.

The experimental data showed the analysis considering to the characteristics: the main significant effects and interactions among the means employed, according to

Araújo et al. 951



**Figure 4.** Stability of the biosurfactant in relation to surface tension free cells in metabolic liquid by *S.marcescens* grown in medium containing cassava wastewater (6%).



**Figure 5.** Response surface of biosurfactant production produced by *S. marcescens* UCP 1549, formulated by the planning factorial  $2^3$  with one central point per block for a total of three blocks determining the surface tension (A - Cassava/lactose); (B – lactose/corn oil); (C – corn oil/cassava).

Factor		Sum of squares (SS)	Degrees of freedom(df)	Mean squares (MS)	F	Р
(1)	Lactose	7.370	1	7.3704	5.6810	0.028372
(2)	Cassava	345.042	1	345.0417	265.9524	0.000000
(3)	Corn oil	476.150	1	476.1504	367.0088	0.000000
1 by 2		55.937	1	55.9371	43.1154	0.000004
1 by3		106.429	1	106.4288	82.0336	0.000000
2 by3		6.912	1	6.9123	5.3279	0.033073
Lack of	fit	146.566	2	73.2831	56.4854	0.000000
Pure er	ror	23.353	18	1.2974		
Total S	S	1167.760	26			

**Table 4.** Analysis of variance of experimental data for the surface tension.

the values F (Fischer constant) and P-(value confidence interval< 0.05). The pure error or experimental error value was considerably reduced; 56.5 times lower than the error on the fit (experimental data).

# Conclusions

The conventional taxonomy and 16S rDNA gene determination confirmed the identification of the pigmented bacterium, as new strain of S. marcescens. In addition, the DNA homology has shown the comparison among all types of prokaryotic microorganisms, and confirmed the identification regardless of their conditions of growth or metabolism and confirmed the identification. The new strain of S. marcescens showed ability to grow on agroindustrial substrate, cassava wastewater and both productions, a potent tensioactive and emulsification agents in the static condition, in the economic and low cost media. The biosurfactant was stable when tested at different pH, temperature and concentrations of NaCl measured by surface tension. The statistic model to tensio-active agent indicated better interactions in the independents variables. The biosurfactant produced by S. marcescens, can be used for bioremediation process of environmental pollution with hydrophobic substances.

# **Conflict of interest**

The authors have not declared any conflict of interest

# ACKNOWLEDGEMENTS

The authors are grateful to National Council for Scientific and Technological Development-CNPq, the Science Foundation and Technology of the State of PernambucoFACEPE, and also to UE GC for the opportunity to develop this research and UNICAP for the use of laboratories.

#### REFERENCES

- Altschul SF, Madden TL, Schäffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ (1997). Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Nucleic Acids Res. 25(17):3389-3402.
- Araújo HWC, Ceballos BSO, Campos-Takaki GM (2009). Biosurfactant production by *Chromobacterium prodigiosum*. In: Current Research Topics In Applied Microbiology And Microbial Biotechnology. pp. 676-680.
- Brumano LP, Soler MF, Silva SS (2016). Recent advances in sustainable production and application of biosurfactants in Brazil and Latin America. Ind. Biotechnol. 12(1):31-39.
- Cirigliano MC, Carman GM (1984). Isolation of a bioemulsifier from *Candida lipolytica*. Appl. Environ. Microbiol. 48(4):747-750.
- Cunha CD, Do Rosario M, Rosado AS, Leite SGF (2004). Serratia sp. SVGG16: a promising biosurfactant producer isolated from tropical soil during growth with ethanol-blended gasoline. Proc. Biochem. 39(12):2277-2282.
- Dundar D, Meric M, Vahaboglu H, Willke A (2009). Pseudo-outbreak of *Serratia marcescens* in a tertiary care hospital. New Microbiol. 32(3):273-276.
- Ebadipour N, Lotfabad TB, Yaghmaei S, RoostaAzad R (2016). Optimization of low-cost biosurfactant production from agricultural residues through response surface methodology. Prep. Biochem. Biotechnol. 46(1):30-38.
- Ferraz C, De Araújo AA, Pastore GM (2002). The influence of vegetable oils on biosurfactant production by *Serratia marcescens*. In: Biotechnology for Fuels and Chemicals. Humana Press. pp. 841-847
- Gorlach-Lira K, Coutinho HD (2007). Population dynamics and extracellular enzymes activity of mesophilic and thermophilic bacteria isolated from semi-arid soil of northeastern Brazil. Braz. J. Microbiol. 38(1):135-141.
- Holt JG, Krieg NR, Sneath PH, Staley JT, Williams ST (1994). Bergey's Manual of determinate bacteriology. pp. 187.
- Joshi SJ, Suthar H, Yadav AK, Hingurao K, Nerurkar A (2012). Occurrence of biosurfactant producing *Bacillus* spp. in diverse habitats. ISRN Biotechnology 2013:1-7.
- Khanafari A, Assadi MM, Fakhr FA (2006). Review of prodigiosin, pigmentation in Serratia marcescens. J. Biol. Sci. 6:1-13.

- Kumer S, Tamura K, Nei M (1993). MEGA: Molecular evolutionary genetics analysis, version 1.0. Pennsylvania State Univ., University Park.
- Kuyukina MS, Ivshina IB, Philp JC, Christofi N, Dunbar SA, Ritchkova MI (2001). Recovery of *Rhodococcus* sp. biosurfactants using methyl tertiary-butyl ether extraction. J. Microbiol. Methods. 46(2):149-156.
- Ma Y, Rajkumar M, Zhang C, Freitas H (2016). Beneficial role of bacterial endophytes in heavy metal phytoremediation. J. Environ. Manage. 174:14-25.
- Montero-Rodríguez D, Andrade RFS, Ribeiro DLR, Rubio-Ribeaux D, Lima RA, Araújo HW, Campos-Takaki GM (2015). Bioremediation of Petroleum Derivative Using Biosurfactant Produced by Serratia marcescens UCP/WFCC 1549 in Low-Cost Medium. Intern. J. Curr. Microbiol. Appl. Sci. 4(7):550-562.
- Olaifa RO, Adeyemi OA, Oloyede ST, Sogunle OM, Agunbiade JA, Okubanjo AO (2015). Performance and carcass characteristics of broiler chickens fed graded levels of cassava peel meal based diets. Malaysian J. Anim. Sci. 18(2):103-112.
- Pessoa GVA, Silva EAM (1972). Meios de Rugai e lisina-motilidade combinados em um só tubo para a identificação presuntiva de enterobactérias. Revits Instuto Adolfo Lutz. 32:97-100.
- Ponte JD (1992). Histórico das pesquisas sobre a utilização da manipueira (extrato líquido das raízes de mandioca) como defensivo agrícola. Fitopatologia Venezuelana. 5(1):2-5.
- Rastegari B, Karbalaei-Heidari HR (2016). Sulfate as a pivotal factor in regulation of *Serratia* sp. strain S2B pigment biosynthesis. Res. Microbiol. 167(8):638-646.
- Rufino RD, Sarubbo LA, Campos-Takaki GM (2007). Enhancement of stability of biosurfactant produced by *Candida lipolytica* using industrial residue as substrate. World J. Microbiol. Biotechnol. 23(5):729-734.
- Sá IB, Cunha TJF, Taura TA, Drumond MA (2015). Mapeamento da desertificação da Região de Desenvolvimento Sertão do São Francisco com base na cobertura vegetal e nas classes de solos. Revista Brasileira de Geografia Física. 8:510-524.

- Saitou, N, Nei M. (1987). The neighbor-joining method: a new method for reconstructing phylogenetic trees. Mol. Biol. Evol. 4(4):406-425.
- Sambrook J, Russell DW (2001). Molecular cloning: a laboratory manual 3<sup>rd</sup> Ed. Coldspring-Harbour Laboratory Press, UK.
- Silva SNRL, Farias CBB, Rufino RD, Luna JM, Sarubbo LA (2010). Glycerol as substrate for the production of biosurfactant by *Pseudomonas aeruginosa* UCP0992. Coll. Surfaces B: Biointerf. 79(1):174-183.
- Solanki RB, Arora S (2015). Leaf litter dynamics in Agroforestry system affecting microbial activity in Saline Soils. J. Soil Water Conserv. 14(4):333-339.
- Sousa Freitas MI, Barbosa JJ, Costa CTF (2015). Uma reflexão sobre mudanças climáticas, saúde e meio ambiente no semiárido nordestino. Saúde e Meio Ambiente: Revista Interdisciplinar. 4(2):61-77.
- Tariq AL, Prabakaran J (2011). Isolation and molecular characterization of UTI associated *Serratia marcescens* TW1 from urine specimens. J. Microbiol. Antimicrob. 3(6):130-135.
- Thompson JD, Higgins DG, Gibson TJ, Clustal W (1984). Improving the sensitivity of progressive multiple alignment through sequence weighting, position-specific gap penalties and weight matrix choice. Nucleic Acids Res. 22(22):4673-4680.
- Vierheilig J, Savio D, Ley RE, Mach RL, Farnleitner AH, Reischer GH (2015). Potential applications of next generation DNA sequencing of 16S rRNA gene amplicons in microbial water quality monitoring. Water Sci. Technol. 72(11):1962-1972.
- Weisburg WG, Barns SM, Pelletier DA, Lane DJ (1991). 16S ribosomal DNA amplification for phylogenetic study. J. Bacteriol. 173(2):697-703.
- Zhu H, Zhou WY, Xu M, Shen YL, Wei DZ (2007). Molecular characterization of *Serratia marcescens* strains by RFLP and sequencing of PCR-amplified 16S rDNA and 16S–23S rDNA intergenic spacer. Lett. Appl. Microbiol. 45(2):174-178.